

American Thoracic Society Documents

ATS Workshop Proceedings: Exhaled Nitric Oxide and Nitric Oxide Oxidative Metabolism in Exhaled Breath Condensate

THIS OFFICIAL WORKSHOP PROCEEDINGS OF THE AMERICAN THORACIC SOCIETY WAS APPROVED BY THE ATS BOARD OF DIRECTORS SEPTEMBER, 2005

CONTENTS

- Background to This Report
- Aims of the Workshop
- Section 1: Update on $F_{E_{NO}}$
 - Background
 - Measurement Issues
- Clinical Application of $F_{E_{NO}}$ Measurement
- Section Summary: Online Measurement of Exhaled NO
- Section 2: Update on Nasal NO Measurement
 - Background
 - The Role of Nasal NO Measurement
 - Conditions in Which Nasal NO Might Be Clinically Applicable
 - How Should Nasal NO Be Measured?
 - Nasal NO and Ostial Patency
- Section Summary: Nasal NO
- Section 3: NO Exchange Models
 - Background
- Section Summary: NO Exchange Models
- Section 4: Exhaled NO in Mechanically Ventilated Patients
 - Background
 - Patient Safety
 - Measurement Techniques and Confounding Factors
 - Pathologic Mechanisms Contributing to Altered $F_{E_{NO}}$ in Critical Illness
- Section Summary: Exhaled NO Measurement in Ventilated Patients
- Section 5: Lung NO and Redox Assessment Using EBC
 - Background
 - Anatomic Source and Potential for Oral Contamination of EBC
 - Issues of Dilution
 - Nitrogen Oxides
 - EBC pH Assays
 - Interactions of pH and NOx
 - Hydrogen Peroxide
 - Evidence of Oxidation by Measurement of Larger Molecules in EBC
- Section Summary: EBC; Redox Status and Nitrogen Oxides
- Overall Summary of Workshop

BACKGROUND TO THIS REPORT

In December 2002, a 2-day American Thoracic Society (ATS)-sponsored workshop was held in Toronto, Ontario, Canada, with the participation of scientists from the United States, Canada,

and Europe working in the relevant field. Several companies who have developed technologies in the field were also present at all scientific sessions, although they were excluded from the session that dealt with revisions to the ATS statement.

The workshop believed that clinical application of exhaled nitric oxide (NO) was ready for asthma, whereas nasal NO could advance to clinical use in primary ciliary dyskinesias. However, exhaled breath condensate (EBC) was not considered ready for clinical application as the now-published recommendations for standardized methods were not ready at the time of the workshop, methodologic issues abounded, and evidence for application of specific markers in specific diseases was deemed inadequate.

AIMS OF THE WORKSHOP

Aim 1: To review developments in the field of fractional concentration of exhaled NO ($F_{E_{NO}}$) measurement.

- New NO sensor technologies, facilitated by Serpil Erzurum
- Online $F_{E_{NO}}$ measurement, facilitated by Philip Silkoff
- Pediatric $F_{E_{NO}}$ measurement, facilitated by Eugenio Baraldi
- Offline $F_{E_{NO}}$ measurement, facilitated by Aaron Deykin
- New concepts in modeling NO output, facilitated by Steven George
- Measurement of $F_{E_{NO}}$ in ventilated patients, facilitated by Nandor Marczin
- Nasal NO measurement and clinical application, facilitated by Jon Lundberg
- $F_{E_{NO}}$ as a clinical tool, facilitated by Philip Silkoff

Aim 2: To review data on the assessment of NO redox status in EBC.

- EBC collection issues, facilitated by John Hunt
- Overview of issues under discussion at the European Respiratory Society (ERS)/ATS task force relating to standardization, facilitated by Ildiko Horvath
- Issues of dilution, anatomic origin, and volatile contribution, facilitated by Richard Effros
- Redox chemistry of nitrogen oxides, facilitated by John Hunt

Aim 3: To update the 1999 ATS statement on exhaled and nasal NO measurement (1) in light of recent developments presented at the workshops.

To highlight how this workshop proceedings document interfaces with the ATS/ERS 2005 statement on exhaled and nasal NO measurement, Aim 3 was realized in that statement (2). The discussions around the bullets in Aim 1 are captured at length in this ATS workshop proceedings document, giving much background to the 2005 updates to the ATS/ERS statement. Aim 2 above, dealing with EBC, overlaps very little with the recently published ATS/ERS task force on EBC, which deals with technical measurement issues and not redox status (3).

An Executive Summary of these Workshop Proceedings is published in the *American Journal of Respiratory and Critical Care Medicine* 2006;173:811-813 (<http://ajrccm.atsjournals.org/cgi/content/full/173/7/811>).

Proc Am Thorac Soc Vol 3, pp 131-145, 2006

DOI: 10.1513/pats.200406-7105T

Internet address: www.atsjournals.org

SECTION 1: UPDATE ON F_{ENO}

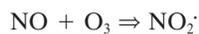
Background

With over 1,000 peer-reviewed publications, research into F_{ENO} is well established, and the time has arrived for clinical application (4). The current online measurement technique is practical, and validated. For clinical use to occur, F_{ENO} monitors must achieve regulatory approval, and procedure reimbursement mechanisms must be developed. In June 2003, the U.S. Food and Drug Administration (FDA) approved the first F_{ENO} analyzer (Aerocrine, Stockholm, Sweden) for monitoring asthma. Technologic advances must allow the production of simple and reliable NO monitors if the cost is to be reduced. The workshop reviewed new sensor technologies, issues of measurement, and the evidence supporting clinical application of F_{ENO} and nasal NO.

Measurement Issues

Sensor technologies.

CHEMILUMINESCENCE. The most widely used approach to NO analysis is chemiluminescence. A number of commercial chemiluminescent NO instruments are available for the measurement of F_{ENO} (Aerocrine [Stockholm, Sweden, and Chicago, IL], Eco Physics and Eco Medics [Ann Arbor, MI, and Berne, Switzerland], Ionics Instruments [Boulder, CO]). Chemiluminescence is based on the reaction between NO in the sample and ozone, which is generated in the analyzer reaction cell as shown in the equations below. The reaction produces emission in the red and near-infrared region of the spectrum ($h\nu$), which is detected by a photomultiplier tube (PMT).



The PMT signal is directly and linearly proportional to the NO concentration in the sample stream.

Newer sensor technologies. As clinical application of NO measurement develops, there will be a need for less complicated technologies and less costly equipment. There are basic requirements for new instruments to be applicable in different settings, such as physician offices and patient homes. These include simplicity, portability, real-time results, suitability for remote analysis, transmission of data, and lack of need to calibrate. The analyzer should, of course, conform to ATS/ERS guidelines. It will be necessary to develop new technology that enables these qualities. Standardization of the measurement and quality control of the results will be key features in equipment intended for primary care. Especially important is to consider facilitation of constant flow exhalation—for example, using dynamic flow control, rejection of nonvalid exhalations, control of ambient NO contamination, and automatic standardization. Analyzer development by several companies is ongoing, but needs to be coordinated with the progress of the methodology in clinical practice. New technologies will increase the availability of NO measurement, which will further reduce the cost as F_{ENO} measurement is incorporated into clinical practice. Eventually, it may be possible to extend F_{ENO} measurement to family practice, and into patients' homes, for self-monitoring.

Other NO analysis capabilities are in development and were briefly presented at the meeting:

EKIPS Technologies (Norman, OK) have employed a tunable diode laser absorption spectroscopy to measure NO and CO₂ in exhaled breath (<http://www.breathmeter.com>). This platform can also measure other gases of interest.

Aerocrine (Stockholm, Sweden, and Chicago, IL) presented a prototype portable NO analyzer, NIOX-Mino

(<http://www.niox-mino.com>), which is based on a new electrochemical sensor technology that measures NO.

Online F_{ENO} measurement updates. The 1999 ATS statement made firm recommendations for online F_{ENO} measurement (1). Certain revisions to this statement were proposed at this workshop for joint ATS/ERS publication, and will be summarized briefly here. These 2005 ATS/ERS recommendations have now been published (2).

Exhalation flow rate. The exhalation rate of 50 ml/s appears to have met with acceptance in both adults and children, although there are only few publications so far that have used this flow rate, or even specified flow rate (5–19). In addition, there is no universal application of this flow rate by professional societies (1, 20) or NO analyzer companies. It is probably acceptable to use different flow rates for research purposes, as long as the flow rate used is described in the study report, and indeed for different diseases, different flow rates may be appropriate. However, for routine clinical application, a single flow rate will be desirable. The plateau concentrations have been reported to be highly reproducible at 50 ml/s compared with higher or lower flow rates in children (14). In addition, at flows above 100 ml/s, no true plateau exists and the tracing is down-sloping (14). Pedroletti and colleagues also found that the discriminatory power of F_{ENO} for asthma was best at 50 ml/s in children (14), although Deykin and coworkers reported that several flow rates between 47 and 250 ml/s had equivalent discriminatory power for diagnosing asthma (6).

Means of controlling flow. When controlling flow, it is possible to have subjects target airway opening pressure (which with a fixed resistance determines a fixed flow) or flow itself (16). Because flow is the prime determinant, it may be better to directly target flow. It is probably adequate that mean flow over a portion of the exhalation falls within a specified range, even if instantaneous flow deviates. There are many different ways to control flow: these include subject targeting of flow or pressure, dynamic or instrument-facilitated flow including dynamic resistors (21), starling resistors (15), and operator-controlled flow (5). These methods are able to compensate for subjects who are unable to precisely control their effort and may be of great use in young children.

Airway pressure. A fixed flow rate can be achieved by using different airway pressures with different resistances. Some investigators have reported that F_{ENO} levels are affected by airway pressure in animals (22), but not in humans (16), but a recent publication suggests the opposite (23). It is possible that in certain airway disease, airway pressure could recruit atelectatic lung, thereby affecting F_{ENO} values.

Exhaled NO plateau definitions. There has been some difficulty in using the plateau definitions as defined by the ATS statement from 1999 (1). The issues are as follows: (1) a first plateau may exist that conforms to the 1999 ATS standards to be followed by a second higher plateau that is probably the real plateau, (2) plateaus are difficult to identify at higher flows as the trace may show a steadily declining NO level (14), and (3) electronic interpretation of plateaus needs overriding by human interpretation on occasions. Plateau definitions need to be practical and easily achievable to avoid having to do an excessive number of exhalations. Although several companies have included software programs that analyze F_{ENO} profiles and identify plateaus that conform to ATS recommendations, visual inspection is also a valid way to determine the plateau. The section on plateau definition has undergone revision and has appeared in the revised 2005 statement (2) as follows:

The duration of exhalation must be sufficient (at least 4 seconds for children < 12 years old and > 6 seconds for children > 12 years

old and adults. This corresponds to an exhaled volume of at least 0.3 L in adults at an exhalation flow rate of 0.05 L/s) to allow the airway compartment to be washed out and a reasonable plateau achieved. In general, patients can exhale comfortably up to 10 seconds and this may be necessary for the achievement of a stable NO plateau. The plateau concentration in NO should be evaluated over 3 seconds (0.15 L) window of the exhalation profile according to the following guidelines. The plateau can be considered to begin at point A and end at point B. The plateau can be flat, positive sloping, or negative sloping. However, the magnitude of the slope should be minimized using the following criteria. Points A and B should be chosen to define the first 3-second window in the exhaled concentration profile such that the absolute magnitude of A–B is less than 10%. In addition, no point within the 3-second window should deviate from either the value at point A or point B by more than 10%. The plateau concentration, $F_{E_{NO}}$, is then defined at the mean concentration over this 3-second window. Once a 3-second plateau is achieved, there is no reason to continue the exhalation.

Number of required exhalations. Many workshop attendees believed that two reproducible $F_{E_{NO}}$ values (agreement within 10% of lower value) are adequate for clinical purposes, although for some research purposes, it may be preferable to continue to require three reproducible values. The revised ATS/ERS statement has included this change (2). If the patient is measuring $F_{E_{NO}}$ for the first time, however, a “training blow” is recommended.

Inhalation phase. The 1999 ATS statement recommends inhalation to total lung capacity (TLC), which is a standard maneuver used in spirometry and is familiar to patients. $F_{E_{NO}}$ levels are reduced when exhaling from lower inspiratory volumes (16). However, inhalation to TLC may be difficult for some subjects. It is now stated in the revised statement that inspiration to TLC is recommended but that inspiration *close* to TLC is acceptable and that subjects should be told to inspire “deeply” or “as deep as you can.”

Calibration gases. There has been concern that calibration gases are in the ppm range, whereas human levels are in the ppb range with assumption of linearity. It has also been difficult to obtain stable, reliable ppb gases in the United States, although these are available in Europe. Eco Physics (Ann Arbor, MI) dilutes a ppm NO gas to the ppb range for calibration. Aerocrine uses a gas of approximately 200 ppb imported from Sweden, and Ionics Instruments uses a 45-ppm gas with linearity tested to the ppb range. Further discussion with industry is necessary to determine feasibility of producing stable ppb gas in the United States. Meanwhile, it is reasonable to continue using current calibration gases.

Normal range of $F_{E_{NO}}$ values. For both research and clinical purposes, it will be necessary to refer to published normal values

in adults and children. There was consensus that such normal data with measures of central tendency and variability should be published in the revised statement or preferably published separately and referenced by the statement. There is a large variability in normal subjects similar to other physiologic parameters.

CLINICAL APPLICATION OF $F_{E_{NO}}$ MEASUREMENT

Asthma

Exhaled NO is well established as a research tool in asthma with over 400 publications regarding this marker. There is no currently recommended periodic assessment of airway inflammation in asthma. The time is ripe for clinical application, as recently reviewed (4). Leaving aside bronchoscopy, which is impractical for routine clinical use, the tests of inflammation available include induced sputum, blood eosinophils and eosinophil cationic protein, urinary leukotrienes, breath condensate, and $F_{E_{NO}}$. Exhaled NO is attractive as the test is completely noninvasive, the cost per test is low, and the result is immediate. This makes it ideal for clinical application. Evidence supporting the clinical utility of $F_{E_{NO}}$ in asthma was reviewed and is summarized in Table 1.

Impediments to the clinical application of $F_{E_{NO}}$ include the paucity of publications regarding the utility of this marker in predicting exacerbations and improving long-term asthma control or reducing health care costs. Although the FDA has cleared one $F_{E_{NO}}$ monitor, reimbursement mechanisms are not yet established. The equipment remains very expensive. Finally, the value of $F_{E_{NO}}$ monitoring is not widely appreciated in the medical community.

Other diseases. Exhaled NO may be a good diagnostic tool for primary ciliary dyskinesia (PCD) syndromes (45), but its clinical utility in chronic obstructive pulmonary disease (COPD), interstitial lung disease, cystic fibrosis (CF), and other pulmonary or extrapulmonary diseases remains to be established.

SECTION SUMMARY: ONLINE MEASUREMENT OF EXHALED NO

This section reviewed some advances in the technologies available for the detection of NO, which will facilitate the progress of this marker from bench to bedside. The 1999 ATS recommendations were reviewed and some changes were recommended to the online exhaled NO measurement technique, which were updated in the revised ATS/ERS statement on exhaled and nasal NO measurement that was published in 2005 (2). Finally, knowledge about the clinical application of exhaled NO was presented and discussed.

TABLE 1. RATING OF EVIDENCE FOR UTILITY OF $F_{E_{NO}}$ IN ASTHMA MANAGEMENT

Utility	Level of Evidence	Potential Application
$F_{E_{NO}}$ raised in asthma (24–29)	5	Diagnosis
$F_{E_{NO}}$ falls after steroids (27, 30–32)	5	Monitoring and titrating response
$F_{E_{NO}}$ falls after other antiinflammatory drugs (e.g., leukotriene antagonists, anti-IgE) (33–35)	2–3	Monitoring and titrating response
$F_{E_{NO}}$ rises on stopping medication (29)	5	Compliance check
$F_{E_{NO}}$ correlates with eosinophilic inflammation (36–39)	2–3	Monitoring the inflammatory component
$F_{E_{NO}}$ correlates with other inflammatory components of asthma (40)	0–1	Monitoring the inflammatory component
$F_{E_{NO}}$ rises with exacerbation (41, 42)	3	Assessment of exacerbation
$F_{E_{NO}}$ predicts exacerbation (43, 44)	1–2	Predicting exacerbation
Targeting therapy to inhibit $F_{E_{NO}}$ to improve control, QOL, health care costs, etc.	0	Improving long-term control
Targeting $F_{E_{NO}}$ to prevent remodeling, FEV ₁ decline, etc.	0	The holy grail

Definition of abbreviation: QOL = quality of life.

For level of evidence, this scale is as follows: 5: > 10 publications; 4: 6–9 publications; 3: 3–5 publications; 2: 1–2 publications; 0: no publications.

SECTION 2: UPDATE ON NASAL NO MEASUREMENT

Background

There is continuous high production of NO in the human nasal passages and paranasal sinuses. Noninvasive measurements of nasal NO are easy to perform and have revealed altered levels in several respiratory disorders. At the 2002 ATS workshop in Toronto, we discussed the latest development in the nasal NO field with focus on two major issues: clinical indications for nasal NO measurements and methodology. In healthy subjects, a large proportion of NO found in exhaled air originates from the upper airways, with only a minor contribution from the lower respiratory tract and the lungs (46–48). The role of NO in the upper airways is not entirely known but may involve host defense functions, including direct toxic effects on microorganisms (49) and regulation of mucociliary function (50). Nasal NO is altered in several respiratory disorders—for example, primary ciliary dyskinesia (PCD) (47), cystic fibrosis (CF) (51, 52), and allergic rhinitis (53–55), and this has led to the proposal that a nasal NO test could be clinically useful in diagnosis and monitoring of these diseases. This section is a brief summary of discussions at the workshop. More extensive reviews of nasal NO are available elsewhere (56, 57).

The Role of Nasal NO Measurement

Nasal NO measurements are rapid, completely noninvasive, and can be performed easily even in infants. Because nasal NO is altered in certain airway disorders, the measurements could be clinically beneficial to aid in the diagnosis and monitoring of therapy. There is a great difference in background NO output between the upper and the lower airways (47, 48, 58). Background lower airway NO output is normally low, which makes it easy to find increases (e.g., asthma) but more difficult to detect decreases. In the upper airways, there is a high background output, and so an increase (e.g., in allergic rhinitis) can be obscured, whereas a decrease is usually easy to reveal. Adding to the complexity is the fact that swelling of the mucosa or secretions during inflammation may lead to less passage of NO from the mucosa of the paranasal sinuses to the nasal cavity where it is measured. In such cases, the net change in nasal NO will be variable and difficult to predict.

Conditions in which Nasal NO Might Be Clinically Applicable

The workshop participants concluded that, as of today, there is one indication for nasal NO measurements that could be considered almost ready to use in the clinic—namely, diagnosis of PCD. Several independent studies have now uniformly reported extremely low nasal NO in PCD. In recent trials, the sensitivity and specificity of this test in PCD has proven to be excellent (59–61). This makes the nasal NO test attractive for screening of PCD, as a guide to confirmatory testing (e.g., ciliary structure analysis). A simple, noninvasive test for PCD could indeed be very useful. Diagnosis of PCD is often delayed despite the presence of typical symptoms early in life (62). The present diagnostic procedure for suspected PCD is quite cumbersome and often involves ultrastructural examination of airway epithelial ciliary structure (63, 64). The use of nasal NO in diagnosis and monitoring of other respiratory disorders (e.g., allergic rhinitis, sinusitis, nasal polyposis, CF) is potentially of great interest, but more research is needed before we know how clinically useful this test can be for these disorders.

How Should Nasal NO Be Measured?

The ATS and ERS have agreed on a highly standardized procedure for measurements of lower respiratory tract F_{ENO} (see section on online NO). Such guidelines are extremely helpful for

researchers when comparing results and they have been of great value in the process of moving F_{ENO} measurements into clinical application. For nasal NO measurements, however, one single standardized measurement procedure has not yet been defined. There are several reasons for this as follows: (1) there has been no well-defined major disease indication (such as asthma for F_{ENO}) where routine measurements of nasal NO can be easily foreseen, (2) the methodology is quite complex, and (3) measurement variability is often substantial. Because of these reasons, the nasal NO field has been driven more by scientific curiosity than by a need for rapid standardization.

The F_{ENO} test for orally exhaled NO is now moving into the clinical arena as a routine test in asthma and will likely also be used when handheld analyzers are available for home use. In parallel, novel, more sophisticated ways of measuring and analyzing F_{ENO} are emerging (see sections on NO models). Such tests add important information and could be particularly useful in specialized centers. A similar development could occur for nasal NO testing. As a first step, it would be desirable to standardize a simple method for a specific disease, and this will most likely be PCD. This would not preclude more sophisticated nasal NO procedures and analyses for other indications.

Several techniques for measuring nasal NO have been used. The most commonly presented way to measure nasal NO is to sample nasal air directly from one nostril. Using the intrinsic sampling flow of a chemiluminescent analyzer or an external pump, air is aspirated from (or insufflated into) one nostril (56, 65). Because of its simplicity, the aspiration method is still by far the most used technique for nasal NO. Another technique to measure nasal NO includes nasal single-breath exhalation using a facemask (66).

If the nasal NO test should become a part of future clinical practice, it will of course be of great importance to standardize the method carefully. However, the recommendation by the workshop at this time was to allow maximum freedom for nasal NO measurements. The chosen method must, however, be carefully described, including reporting of flow rate and ambient NO levels. It is likely that several methods for nasal NO will be used depending on the situation and the patient population. For example, breath-holding or single-breath measurements are not possible in noncooperating individuals (e.g., infants and sedated patients).

Nasal NO and Ostial Patency

One of the more recent findings in this field is the intriguing discovery that nasal NO increases dramatically (5- to 15-fold) during humming compared with silent nasal exhalation (67). The oscillating sound waves may speed up the exchange of gases over the sinus ostium, resulting in a rapid washout of NO from the sinuses (68). This NO peak is transient and the levels decrease gradually during repeated humming maneuvers. However, nasal NO levels fully recover after a short period of silence, which allows sinus NO to accumulate again. In a model of the nose and sinus, it was found that ostium size was the main determinant of the humming-induced increase in NO. In addition, in patients with computed tomography-proven complete sinus ostial obstruction (bilateral nasal polyposis), the nasal NO increase during humming was abolished (69). This suggests that the increase in nasal NO during humming correlates to ostial function. Ostial obstruction is central in the pathogenesis of sinusitis, and one goal in medical and surgical therapy of chronic sinusitis is to improve sinus ventilation. Future studies will show if a nasal NO humming test could be used to monitor ostial function in at-risk patients or in subjects with established sinus disease.

SECTION SUMMARY: NASAL NO

About a decade ago, it was found that NO is released in large quantities in the nasal passages of healthy humans. Especially large concentrations of NO have been found in the paranasal sinuses. The physiologic role of this NO still has not been clarified but may include important local host-defense mechanisms. Nasal NO can be measured online using different noninvasive techniques. Using these methods it has been found that nasal NO is altered in several airway disorders, including allergic rhinitis, PCD, CF, and sinusitis. For most indications, nasal NO is still to be regarded as an interesting research tool with potential clinical importance. However, in PCD, the situation is different because nasal NO is uniformly extremely low. The sensitivity and specificity of a nasal NO test in PCD are so good that this test now should be considered for routine use in specialized centers. To facilitate this process, we need to quickly establish guidelines for a simple, standardized nasal NO test in PCD.

SECTION 3: NO EXCHANGE MODELS

Background

NO was first discovered in exhaled breath of humans and other vertebrates in 1991 (70). This finding spurred an era of intense investigation aimed at determining the source and potential use of this noninvasive biological signal. In 1997, the concentration of NO in the exhaled breath was reported to be highly dependent on the exhalation flow rate (71, 72), differing significantly from other endogenously produced gases such as carbon dioxide and nitrogen. This finding necessitated the development of new models and analytic methods to understand the underlying physiology and gas exchange mechanisms.

A simplified two-compartment model of the lungs was presented in 1998 (73), which explained many of the unique features of NO exchange dynamics—in particular, the dependence on exhalation flow rate. The two-compartment model describes $F_{E_{NO}}$ as having significant sources from two compartments—namely, the airways and the alveolar region. Thus, NO exchange dynamics are described using three flow-independent exchange parameters. One describes the alveolar region: $C_{A_{NO}}$, the steady-state alveolar concentration; and two describe the airway region: (1) the airway NO diffusing capacity ($D_{aw_{NO}}$), alternatively described as the conductance for mass transfer or transfer factor of NO between the airway tissue and the gas phase, and (2) the maximum airway wall flux ($J'_{aw_{NO}}$); $J'_{aw_{NO}}$ is equal to $D_{aw_{NO}} \times C_{aw_{NO}}$ (the airway wall NO concentration).

Since the original description of the two-compartment model, research has focused in three areas: (1) developing experimental breathing and analytic techniques to accurately and reproducibly estimate the flow-independent NO exchange parameters, (2) estimating the flow-independent NO parameters in health and disease, and (3) further development and testing of the underlying assumptions of the simple two-compartment model. Together, these research thrusts have enhanced our understanding of NO exchange mechanisms and the pathophysiologic interpretation of the flow-independent NO exchange parameters.

The two-compartment model has been previously described in detail (73–76). Figure 1 depicts the basic features of the model. NO is produced by cells in the alveolar membrane producing a net flux of NO into the alveolar space. During an exhalation or breath-hold of more than approximately 8 to 10 s (73, 77–81), the concentration in the alveolar region, $C_{A_{NO}}$ (ppb), reaches a steady state. As alveolar air is convected through the airways toward the mouth during exhalation, additional NO is absorbed by the gas stream from the airway walls denoted by the flux of NO from the airways, $J_{aw_{NO}}$ (rate of NO transferred per unit

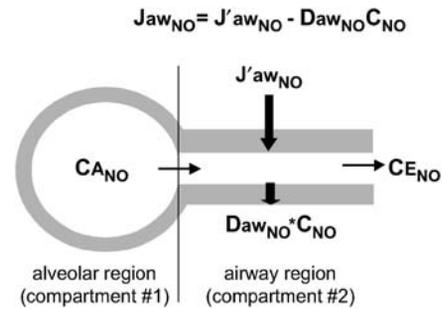


Figure 1. Schematic of two-compartment model used to describe nitric oxide (NO) exchange dynamics. Exhaled NO concentration, $C_{E_{NO}}$, is the sum of two contributions—the alveolar region and the airway region—which depend on three flow-independent parameters: maximum total volumetric flux of NO from the airway wall ($J'_{aw_{NO}}$, picoliters [pl] \cdot s^{-1}), diffusing capacity of NO in the airways ($D_{aw_{NO}}$, $pl \cdot s^{-1} \cdot$ part/billion), and steady-state alveolar concentration ($C_{A_{NO}}$, part/billion). $J_{aw_{NO}}$ is the total flux (pl/s) of NO between the tissue and gas phase in the airway, and is an inverse function of the exhalation flow rate, \dot{V}_E .

time in picoliters [pl]/s). $J_{aw_{NO}}$ is expressed as a linear function of the airway gas phase NO concentration, C_{NO} , by the following equation (73):

$$J_{aw_{NO}} = J'_{aw_{NO}} - D_{aw_{NO}}C_{NO}$$

or

$$J_{aw_{NO}} = D_{aw_{NO}}(C_{aw_{NO}} - C_{NO})$$

$J'_{aw_{NO}}$ is the maximum flux of NO from the airway tissue that is equal to the airway compartment flux if airway gas phase NO, C_{NO} , was zero occurring at infinite exhalation flow, or alternatively, simply the product $D_{aw_{NO}} \cdot C_{aw_{NO}}$.

Once the flow-independent parameters are known, the two-compartment model can be used to predict the exhaled concentration of NO, $C_{E_{NO}}$ (also represented symbolically as $F_{E_{NO}}$), at any constant exhalation flow using the relatively simple exponential expression:

$$C_{E_{NO}} = C_{aw_{NO}} + (C_{A_{NO}} - C_{aw_{NO}}) \cdot \exp(-D_{aw_{NO}}/\dot{V}_E)$$

where \dot{V}_E is the constant exhalation flow rate.

The potential advantage of the flow-independent NO parameters lies in their ability to partition $F_{E_{NO}}$ into two important anatomic subdivisions of the lungs—namely, the airways and the alveolar region, each with their own pathologic involvement. To uniquely determine all three flow-independent parameters, both low (< 50 ml/s) and high (> 100 ml/s) exhalation flow rates must be sampled (74–76, 82). Three approaches have been described in the literature: multiple constant exhalation flows (73–76, 83), a dynamically changing flow within a single exhalation (82), and tidal breathing methods (84). Recently, the impact of axial diffusion of NO toward the alveolar region during exhalation on the estimated parameters has been described (85–87).

The flow-independent NO exchange parameters have been estimated in healthy adults, children, and infants, as well as in several diseases, including asthma (76, 88–93), allergic alveolitis (89, 91), CF (15), scleroderma (94), allergic rhinitis (88), and COPD (88), and in smoking (88, 95). Although asthma is a heterogeneous disease, it appears that steroid-treated subjects with asthma have an elevated $D_{aw_{NO}}$, whereas subjects with asthma who are not treated with steroids have elevated $D_{aw_{NO}}$, $J'_{aw_{NO}}$, and $C_{A_{NO}}$. Subjects with allergic alveolitis have elevated $C_{A_{NO}}$, whereas subjects with scleroderma have an elevated $C_{A_{NO}}$ but a reduced $J'_{aw_{NO}}$. Subjects with CF have an elevated $D_{aw_{NO}}$

but a reduced C_{awNO} . Allergic rhinitis appears to increase D_{awNO} . For subjects with COPD, the C_{ANO} is elevated, and in smoking, C_{awNO} and J'_{awNO} are both reduced.

SECTION SUMMARY: NO EXCHANGE MODELS

Exhaled NO has significant sources from both the airway and alveolar regions of the lungs. This feature has stimulated the development of new analytic techniques to understand this rich feature. To date, a two-compartment (airways and alveoli) model of the lungs has proven to be a simple and robust means of describing NO exchange dynamics. The two compartments are described by three flow-independent NO parameters, two of which characterize the airway compartment (D_{awNO} and C_{awNO}) and one which characterizes the alveolar compartment (C_{ANO}). Several analytic techniques have been used to estimate these parameters in both health and disease. Early reports suggest that the flow-independent NO parameters are uniquely altered in several inflammatory disease states and thus may provide pathophysiologic insight or assist in the clinical management of lung diseases. Future studies must address several important model simplifications, as well as improve our understanding of the clinical and physiologic significance of alterations in these parameters.

SECTION 4: EXHALED NO IN MECHANICALLY VENTILATED PATIENTS

Background

The anesthetic and critical care community has expertise with breath analysis; e.g., capnography and anesthetic gas monitoring during patient care. Extension of this expertise to exhaled breath markers of inflammation would provide this community valuable insight into metabolic alterations that potentially contribute to critical illness (96). Thus, parallel to increased understanding of the role of breath markers in spontaneously breathing patients, a number of marker molecules have been identified in the breath of mechanically ventilated patients that may be useful in identification of disease progression, or in monitoring therapeutic efficacy (97–102).

Despite this progress, the Toronto workshop participants believed that there was insufficient knowledge to make firm recommendations for guidelines for F_{ENO} measurement in ventilated patients for inclusion in the revised ATS/ERS statement on exhaled and nasal NO measurement. Therefore, the session on F_{ENO} in mechanical ventilation was used to overview available information on determinants of F_{ENO} and published measurement approaches.

Patient Safety

The ATS group stressed the need to address patient safety when measuring F_{ENO} for research purposes. Although outpatient measurement of F_{ENO} is considered a low-risk research procedure, the same in ventilated, critically ill patients involving alterations in the ventilation circuit, ventilator settings, and inhaled gases requires special consideration in the unique environment of the operating room or intensive care unit, where electrical hazards and inflammatory gases exist. Where measurement involves interrupting ventilation and disconnecting the ventilator circuit, caution is recommended not to use excessive inflation volumes, to prevent inadvertent hypoxia and infection, and accidental extubation.

Measurement Techniques and Confounding Factors

Several approaches have been developed for online NO testing, including tidal, mixed expired, and flow-controlled single-breath analysis.

Breath-to-breath (tidal) F_{ENO} measurements. Fast-response chemiluminescence analyzers are now able to resolve the NO concentration profile within one respiratory cycle, while ventilation is standardized over a measurement epoch. Breath-to-breath analysis results in a dynamically changing NO concentration versus time plot, reflecting NO production in the lower airways of intubated patients. It is likely that this pattern is related to continuously changing expiratory flow. Simultaneous recording of CO_2 concentration and flow profile greatly facilitates data analysis and interpretation where sample-tubing characteristics, lag phase of NO, CO_2 and flow measurements, and variable response times of each of these analyzers should be considered.

The NO versus time trace obtained by breath-to-breath analysis can be described in several ways, including the following: (1) the peak F_{ENO} concentration, (2) mean F_{ENO} , (3) the area under the concentration curve for an indicated time period, or (4) NO output rate \dot{V}_{NO} (nl/min) (103).

The anatomic origin of F_{ENO} has great implications for data interpretation and the utility of this marker to monitor alveolar inflammation in critically ill patients. The participants agreed that very low alveolar NO concentrations (104–106) cause signal-to-noise ratio issues, and disease states causing a decrease in alveolar NO concentrations will likely face problems of lower detection limit (104, 107). In contrast, mechanisms that increase pulmonary microvascular and alveolar NO, such as endothelium-dependent vasodilators and NO donor drugs, appear to produce a readily detectable increase in F_{ENO} (108–112), which might be useful in probing this important compartment.

DETERMINANTS OF BREATH-TO-BREATH F_{ENO} PROFILE. Animal and human studies indicate four major categories of determinants: (1) composition of inhaled gases, (2) ventilation parameters during controlled mechanical ventilation, (3) different ventilation modes, (4) pulmonary blood flow and intrathoracic blood volume.

Among inhaled gases, NO, O_2 , and CO_2 should be carefully considered (106, 113–116). Within a single ventilation mode, F_{ENO} concentration in ventilated patients is affected by respiratory rate, tidal volume, and inspiratory/expiratory ratio, even when minute ventilation is held constant (104, 117). In addition, bias flow from the ventilator and the influence of positive end-expiratory pressure require special attention (118–120). It is also evident that different ventilation modes, such as pressure-controlled inverse ratio ventilation and high-frequency oscillatory ventilation, influence F_{ENO} through multiple mechanisms (121, 122). It has been shown that significant acute reduction in perfusion pressure, such as seen during hemorrhage, onset of cardiopulmonary bypass, occlusion of pulmonary vessels, and correction of left to right intracardiac shunts, alters F_{ENO} (107, 120, 123–126). However, moderate changes in either direction from normal blood flows do not alter F_{ENO} concentrations significantly (119, 127).

Breath-holding. This simple maneuver might be quite useful in some ventilated patients, especially when there are limitations of the tidal breath measurements. Gaseous NO exhibits exponential increase during breath-holding and reaches a plateau phase within 20 to 30 s (104, 106, 128). Because the plateau phase can be explained by a steady state between NO production/release to the gas phase, and consumption/removal from this phase presumably by pulmonary blood flow, different measures of F_{ENO} during breath-holding (slope and plateau phase) might provide useful information regarding NO production and consumption in the lung without the confounding effect of different ventilation parameters and modes.

Real-time analysis of mixed-expired gas. This alternative method relies on the measurement of \dot{V}_{NO} , the rate of elimination

TABLE 2. COMPARISON BETWEEN ONLINE AND OFFLINE NITRIC OXIDE MEASUREMENTS IN MECHANICAL VENTILATION

Online	Offline
<p>Advantages</p> <ul style="list-style-type: none"> Suboptimal measurements can be immediately identified and discarded or repeated Allows for simultaneous measurement of flow Allows precise timing of breaths NO levels can be determined at different points in the respiratory cycle <p>Disadvantages</p> <ul style="list-style-type: none"> Requires more stringent analyzer specifications Patient and analyzer need to be at the same place at the same time Less efficient use of analyzer (more analyzer time per patient) 	<ul style="list-style-type: none"> Samples can be collected from sites remote from the analyzer Less dependent on analyzer response times More efficient use of analyzer (less analyzer time per patient) Allows for measurement of other gases in the same sample <ul style="list-style-type: none"> Possible errors due to sample storage and transport Suboptimal measurements cannot be immediately identified More likely to be affected by ventilator variables Provides NO level in a mixed-gas sample and not at a defined point in the respiratory cycle.

Definition of abbreviation: NO = nitric oxide.

of NO from the lungs. A mixing chamber is connected to the exhaust port of a ventilator, and a real-time NO analyzer is used to sample NO concentration in the gas exiting the mixing chamber. The minute ventilation is measured by means of an online spirometer connected with the mixing chamber. After standard adjustments for ambient temperature and pressure, \dot{V}_{NO} is the product of the 1-min volume entering the chamber and the concentration in the chamber. This method may be used with any mode of ventilation.

Flow-controlled single-breath measurements. To obtain flow-independent parameters in intubated, nonparticipating patients, a method for multiple, single-breath exhalations at various flow rates has recently been proposed (128). This requires disconnection from the ventilator circuit and performing a manual deep inspiration followed by a flow-controlled exhalation using an aspiration suction device. This method allows for repeated measurements at various controlled exhalation flow rates, both in the intubated and the awake state of the same patient. A potential disadvantage might be alveolar collapse during disconnection from the respiratory circuit. Caution is recommended not to use excessive inflation volumes, to prevent inadvertent hypoxia, accidental extubation, and infection.

Offline methods. Here, exhaled gas is collected into an inert reservoir for detection at a different time or location. Offline techniques offer several added advantages compared with online measurements, including the following: (1) portability, (2) less dependency on analyzer response times, (3) more efficient use of analyzers, and (4) potential to measure other gases in the same sample (see Table 2) (129).

At the time of the workshop, there was limited published information on the offline measurement of $F_{E_{NO}}$ levels in ventilated individuals (103, 125, 126, 130, 131). These studies used various methods of gas collection (syringe aspiration, tidal breath collection, and controlled collection with a pump) and different ports of sampling (endotracheal tube, ventilator circuit, and exhaust port of the ventilator). Furthermore, single or multiple breaths were collected for analysis. It appears that variability of NO levels obtained by these different methods was very high, which underscores the need to standardize offline measurement issues.

Pathologic Mechanisms Contributing to Altered $F_{E_{NO}}$ in Critical Illness

Activation of the constitutively expressed NO synthase (NOS) isoforms, induction of type II NOS, and production of NO from acidified nitrite are the principal mechanisms contributing to increased NO production in the lung and potentially causing higher than normal $F_{E_{NO}}$ (132–134). The main mechanisms that would decrease NO bioavailability and concentrations include

down-regulation of constitutive NOS expression, inability to induce type II NOS by inflammatory cytokines and consumption reactions of NO in the fluid phase. (135, 136).

Clinical Implications of Exhaled NO in Critical Care Medicine

Exhaled NO is elevated in various forms of pulmonary infection, including intensive care patients with developing ventilator-associated pneumonia and in lung transplant recipients with infective complications (137).

The characteristics of $F_{E_{NO}}$ in sepsis remain controversial, possibly due to species differences (138–140). In healthy volunteers, lipopolysaccharide administration produced only a modest increase in $F_{E_{NO}}$ (141). Little is known regarding $F_{E_{NO}}$ in patients with clinical sepsis, severe sepsis, or septic shock.

There are also species differences regarding $F_{E_{NO}}$ in acute lung injury (142, 143). In patients with established clinical adult respiratory distress syndrome, levels of $F_{E_{NO}}$ were decreased (144). Further studies are needed to delineate kinetics of gaseous and fluid phase levels of NO during the entire spectrum of the disease (145).

The relationship between $F_{E_{NO}}$ and lung injury associated with ischemia–reperfusion during cardiac surgery and transplantation has received particular attention. Dysfunction of NO pathways and reduced $F_{E_{NO}}$ have been demonstrated in children after cardiopulmonary bypass (125, 130). In adults, $F_{E_{NO}}$ was either increased or decreased or unaltered after cardiac surgery (103, 131, 146–148). In conditions of more severe lung ischemia and reperfusion, such as during lung transplantation, $F_{E_{NO}}$ was reduced in the majority of patients, which appeared to correlate with clinical outcome (149).

SECTION SUMMARY: EXHALED NO MEASUREMENT IN VENTILATED PATIENTS

In summary, the Toronto ATS meeting was an important step in our worldwide dialog toward international consensus and guidelines on measurement issues and on the value of $F_{E_{NO}}$ in critical illness. However, further studies are needed to clarify many of these initial suggestions before the technology can be considered as a diagnostic tool of inflammation in ventilated, critically ill patients.

SECTION 5: LUNG NITROGEN OXIDE AND REDOX ASSESSMENT USING EBC

Background

There are two principal discrete purposes for studying EBC. First is that this fluid can safely provide information about airway lining fluid (ALF) composition. Second is the potential for EBC

assays to provide evidence of airway disease, particularly in regard to inflammation and redox disturbance.

Many of the early studies of EBC related to redox monitoring. Hydrogen peroxide and nitrogen oxides have been studied more than other compounds. Because acidity is a critical determinant of many redox activities (chemical reactivities and enzyme function), EBC pH has also received attention. This workshop focused on the potential of EBC to provide information regarding the otherwise difficult to assess lung NO and redox balance. Studies of exhaled cytokines, leukotrienes, and other compounds of interest in EBC were left to other forums. Summaries of data presented and discussions held are presented below.

Anatomic Source and Potential for Oral Contamination of EBC

In contrast to endotracheal collections, when collecting samples of EBC orally there is likely some contribution from the upper airway. The oropharyngeal secretions contain substantial concentrations of many of the biochemical markers that have been identified in EBC, and slight contamination of a sample with oral secretions has potential to greatly affect EBC concentrations. The various EBC collection systems used by laboratories usually incorporate methods to limit gross salivary contamination.

Theoretically, regions of turbulent airflow, such as carinae and cartilage rings, should provide more particles to the exhaled airstream, although airway reopening after closure may aerosolize particles. The source of the gas phase (volatile) constituents is more likely considered the entire airway and the alveoli.

Issues of Dilution

To gain insight into precise concentrations of nonvolatile constituents of ALF, it is necessary to determine to what extent the particles of ALF are diluted by condensed water vapor. Several methodologies for this are becoming available.

When the substances of interest are volatile, the dilution issue becomes entirely different and in many ways irrelevant.

Importantly, just as has always been accepted for cellular comparisons in bronchoalveolar lavage fluid (BALF), ratios of substances in EBC can be enlightening without need for dilution markers. Ratios of nitrite (NO_2^-) to total nitrogen oxides (NOx), acids to bases (pH), oxidized glutathione (GSSG) to reduced glutathione (GSH), and others are therefore likely to be of interest. Of course, as for BALF, whether one compound is rising or the other is declining will not necessarily be clear.

Nitrogen Oxides

Multiple studies have identified differences in EBC NOx concentrations in various lung diseases. Initially undertaken in an effort to develop a surrogate assay for the more expensive F_{ENO} assay (150), it became clear that the complexities of nitrogen oxide chemistry make EBC NOx assays and F_{ENO} complementary, not equivalent.

Although NO in the airway can be formed by the NOS enzymes (151, 152) and released as gas in the exhaled air, only a small portion of NO thus formed is released. Some is oxidized to become reactive species with downstream signaling effects. Species such as peroxynitrite and peroxynitrous acid may then lead directly to nitration reactions forming nitrotyrosine. Some NO becomes incorporated in various S-nitrosothiols, which serve as storage molecules for NO activity, and have key signaling properties of their own, discrete from NO (153). Much NO is oxidized to NO_2^- and NO_3^- . These less reactive species may be reabsorbed by the airway, carried up the airway in the ALF and swallowed, or exhaled in particles. Nitrite may be consumed by eosinophil peroxidase (154) and neutrophil myeloperoxidase as substrate for enzymatic nitration. When protonated, nitrite will

release NO (155). Likewise S-nitrosothiols also release NO (156). Bacterial and fungal enzymes can reduce NO_3^- , NO_2^- , and NO, reversing the normal eukaryotic pathway, and on complete reduction, form ammonia (157). Many of the intermediaries and final products listed above are found in EBC.

Lung NOx, including F_{ENO} , are best understood when considered as a group, and then especially if other chemical species are considered, such as superoxide and hydrogen peroxide. The bacterial load and the pH of the airway environment need to be considered. Inhalation or endogenous formation of oxidants may affect F_{ENO} , in the same manner that inhaled NO may alter exhaled H_2O_2 (158). Deviations of EBC NOx concentrations, or ratios among them, may reflect the oxidative and nitrogen oxide conditions of the airway, including not just formation of NO, but all the various inorganic, eukaryotic, and prokaryotic reactions that occur around NO. These assays then may reflect NOS activity, airway inflammation, innate immune responses, bacterial burdens, airway pH, and nitrgergic neurotransmission.

Additional values of NOx assays include the potential for them to provide information regarding ventilator-induced lung injury and acute respiratory distress syndrome (ARDS) (159, 160). In this regard, Drs. Gessner and Wirtz presented data at this workshop from their studies of acute lung injury in humans, identifying strong correlations between EBC NO_2^- concentration and the tidal volumes used for ventilation. The ratio of NO_2^- to tidal volume correlated highly with injury severity scores. Nitrite may be formed in association with mechanical stress, with overexpansion of open respiratory units leading to increasingly high and readily measurable EBC NO_2^- output (159). This simple assay may allow titration of ventilatory volumes while monitoring objective evidence of evolving lung injury hour to hour.

The biochemical source of aqueous phase nitrogen oxides in EBC remains obscure. Nitrate and NO_2^- in EBC may be present because these ions are in the particles of ALF. Or they could occur from exhaled gas phase NO being oxidized *ex vivo* in the EBC (e.g., by hydrogen peroxide). Thus, NOx may need to be considered in part as indirectly volatile constituents of EBC.

Cautious techniques need to be used for EBC collection, storage, and assay when NO_2^- or NO_3^- are the ions of interest. These NOx are ubiquitous laboratory contaminants, abundant on fingertips that are carried in humidity and deposit on every available surface. Protection by covering lab surfaces is not sufficient because NO_2^- and NO_3^- are also formed from NO oxidation, and NO gas is in almost all labs and clinics at variable, but relevant, concentrations. NO can travel through plastic coverings as well. Latex gloves may have extremely high levels of NOx on their surface. NOx almost always substantially contaminates microcentrifuge tubes and other test tubes. This is an important issue, particularly for NO_2^- , because it is found in low to submicromolar ranges in EBC, and thus contamination, though unavoidable, can overwhelm the signal from the subject. In addition, NO_2^- can be oxidized to nitrate in aqueous solution, or be converted to NO (through protonation and release of NO from decomposed nitrous acid). These processes can decrease nitrite levels during storage over time.

EBC pH Assays

Most redox reactions are very sensitive to the local pH. Acids tend to be volatile from acidic fluids, and bases volatile from basic solutions. One theoretic underpinning for studying EBC pH is that volatile airway acids could be trapped in EBC, and that they would be exhaled to a greater extent from an acidic source fluid. Thus, a low EBC pH would reflect a low ALF pH, which in turn would influence redox chemistry. Relative to controls, the pH of EBC has been found to be significantly

low in diverse respiratory diseases, including asthma, COPD, bronchiectasis (161, 162), CF (163), and exacerbations of these diseases, as well as in ARDS (164). Correlations of pH are present with relevant cellular inflammation in induced sputum, EBC cytokines, nitrogen oxides, hydrogen peroxide, and 8-isoprostane (161, 164).

The pH of EBC can be measured before or after gas standardization. Gas standardization involves bubbling a CO₂-free gas through the EBC sample for several minutes. Gas standardization removes carbon dioxide and bicarbonate from the EBC (although complete removal is not certain), and the pH rises and then stabilizes during the process. Without gas standardization, the EBC pH is affected by ambient and exhaled carbon dioxide and is not stable, and therefore should be measured immediately on collection. Gas standardization is useful when samples cannot be assayed immediately, when one is not interested in exhaled carbon dioxide levels, or when the EBC pH is expected to be particularly acidic (below a pH of 5, removal of carbon dioxide has little effect on pH, consistent with pH being a logarithmic scale).

Gastroesophageal reflux of acidic fluid, with or without microaspiration into the trachea, is one potential mechanism of EBC acidification that has been considered and is under active investigation in several laboratories. Alternate explanations for EBC acidification include lower airway acid production by numerous described pathways (165). In likelihood, any process that acidifies any level of the airway could lead to EBC acidification if anions present in the fluid become protonated and volatilized.

Ammonia concentrations in EBC range up to 3 mM (166), derived predominantly, but not completely, from the upper airway (167, 168). It has been noted that a low EBC pH is always accompanied by low EBC ammonia concentration, although the obverse is not true. There are reports of isolated lower airway samples of EBC being acidic in various diseases, but not in healthy controls. Although there is general consensus regarding the utility of EBC pH as a simple marker of intuitive value in airway disease, this consensus is not unanimous, as alternative explanations for EBC acidification other than lower airway pH deviation have been presented. There are reasoned theoretic concerns that ammonia derived from the mouth might interfere with EBC pH assays (169), but these are not borne out as yet by empiric investigation. A recent study collecting EBC from a cuffed endotracheal tube from 32 subjects, altogether bypassing the mouth and its ammonia contribution, revealed EBC pH values unchanged from same-subject oral collections (170), despite the expected substantial declines in the levels of ammonia (168). Although this provides support to the concept that EBC pH is a valid measure of lower airway pH, also conceivable is the notion that bypassing the mouth may prevent oral acid(s) from entering the EBC, which could lead to the pH from the isolated lower airway samples being coincidentally the same as the oral collections. Data will assuredly emerge over time that will clarify these issues.

It will help to have more invasive pH measurements of the lower airway during active disease states to correlate EBC pH with airway pH deviations. However, it is possible that EBC acids may be volatilized from airway sources too low to access with other methods, or that discrete small areas of substantially low airway pH may lead to sufficient volatilization of acids to acidify the EBC. Extensive studies of EBC pH in the isolated lower airway of endotracheally intubated patients are ongoing and should clarify the anatomic source and chemical and clinical relevance of this noninvasive effort to gain an understanding of lower airway chemistry.

Interactions of pH and NOx

In animal models (171), experimental airway acidification leads to production of NO measurable in exhaled breath, and the

speed with which this process occurs after instillation of acid (seconds) is consistent with inorganic decomposition of protonated nitrite (nitrous acid = HNO₂) forming, in the end, NO. This has been proposed to occur in acute asthma, where the most profound declines in EBC pH have been noted (162). Also, breakdown of s-nitrosoglutathione leads to increased exhalation of NO (172). It seems likely that there are multiple pathways of NO formation in addition to up-regulation of NOS isoforms, each with relevance to certain conditions.

Dr. Redington examined the relationship between EBC pH and F_{E_{NO}} in patients with mild, stable allergic asthma, patients with stable CF, and control subjects. F_{E_{NO}} was measured online at a flow rate of 250 ml/s and non-gas standardized EBC pH was measured immediately after collection. These findings demonstrate dissociation between non-gas standardized EBC pH and F_{E_{NO}} in these conditions (173, 174). The chemistry of the NOx remains complex. Oxidative burdens in CF may lead to more NO₃⁻ and less NO₂⁻ formation from NO, thus trapping NO, while limiting NO₂⁻ sources for inorganic NO formation.

Hydrogen Peroxide

One of the first markers found in EBC in various disease states was H₂O₂. Several assays have been used, and normative values for children have been published (175). As with other chemical processes in the lower respiratory tract, *in vivo* assessment of the importance of H₂O₂ in disease states had been nearly impossible until the advent of EBC assays. Indeed, EBC has provided the strongest evidence to date of oxidative disturbance in lung diseases.

However, concerns about H₂O₂ flow dependence (176) and chemical reactivity support efforts to standardize EBC collection and assay techniques when this compound is of interest. Similar to F_{E_{NO}}, evidence exists that lower controlled (nontidal breathing) exhalation flows lead to significantly higher EBC H₂O₂ levels than higher controlled flows. Levels of H₂O₂ in EBC decline over time in storage. It is unclear how rapidly H₂O₂ is consumed in EBC, although it is likely that some is consumed even during the time of collection. Hydrogen peroxide will react with NOx, and can react with ammonia also, forming various compounds, including hydroxylamine, peroxyxynitrite, and NO₃⁻. To simplify understanding of H₂O₂ in the airway, the development of online measurement techniques may prove valuable as a standardized assay technique. In this regard, Dr. Becher presented data on a new instrument designed to provide rapid H₂O₂ assay immediately on collection of sample. The instrument, designed and manufactured by FILT in Germany, uses a biosensor for assaying small volumes of EBC, and completes the assay within several minutes.

The issue of flow dependence for EBC H₂O₂ is very different than for NO, as tidal breathing during EBC collection incorporates multiple different flows in each breath. Standardizing to many minutes of consistent-flow controlled exhalation may not be reasonable to demand from a patient.

An alternative to performing the H₂O₂ assay immediately has been presented. This involves addition of a reagent to the just-collected EBC (177), or to the collection device in advance of collection. This reagent reacts rapidly and specifically with hydrogen peroxide and allows for delayed spectrophotometric analysis, as much as 24 hours subsequently.

H₂O₂ is volatile, although less so than water, and the concentration in EBC may be derived both from gaseous H₂O₂ as well as ALF particles. The relative contribution of each source remains incompletely understood.

Evidence of Oxidation by Measurement of Larger Molecules in EBC

Dr. Corradi and colleagues presented data from their investigations of biomarkers of lipid peroxidation (aldehydes and 8-isoprostane) and an antioxidant (glutathione, GSH) in the EBC of children with asthma during acute asthma exacerbation and after 5 days of therapy with prednisone (178). Aldehydes and GSH were analyzed with liquid chromatography and detected with the most sensitive detection systems available today—namely, fluorescence detection and mass spectrometry techniques. 8-Isoprostane was measured by enzyme immunoassay. The data showed that the levels of malondialdehyde and 8-isoprostane were significantly higher in EBC of children with acute asthma than in control children. On the contrary, the levels of GSH were significantly lower in children with acute asthma compared with healthy control subjects. After 5 days of systemic corticosteroid treatment, GSH levels rose, whereas malondialdehyde and 8-isoprostane fell. After oral prednisone therapy, 8-isoprostane levels remained higher in children with asthma than in healthy children, suggesting that corticosteroids may not be fully effective in reducing oxidative stress during acute asthma exacerbation.

The relative changes in the reduction/oxidation chemistry in biomarker levels during acute asthma (highly turbulent airflow) and after treatment with systemic corticosteroids (reducing airway obstruction and turbulent airflow) support that changes in the concentration of biomarkers in EBC samples obtained from subjects with airflow limitation do not purely reflect increased aerosolization of ALF particles. Indeed, the biochemical changes in exhaled breath parallel current theory regarding redox disturbance in the lung, and their measurement now serves as a means to allow such theories to be tested.

Standardization Issues

There are currently two commercial EBC collection systems available, plus numerous homemade systems for sampling EBC. The various devices and techniques likely differ in many respects in regard to condenser materials, temperature of collection, efficiency of condensing water vapor, and efficiency of collecting ALF droplets. There may be temperature-dependent issues that affect the ability of different devices to trap volatile substances. Because EBC is collected from mixed air arising partially from deep airways but also from anatomic dead space and upper airways, breathing pattern, frequency, tidal volume, inspiratory breath holding, and minute ventilation may influence constituents. These issues need to be considered when interpreting EBC concentrations of redox-relevant substances. Standardization of collecting techniques and assays may assist in interlaboratory data sharing. However, the various substances in EBC have different reactivities with other EBC constituents, and differing stabilities. Thus, it would appear unlikely that one collection technique will be optimal for all compounds of interest in EBC. Some investigators urge standardization now. Others suggest that standardization too early may inhibit the development of newer and more effective techniques for studying exhaled breath constituents.

In this regard, Drs. Gessner and Wirtz presented data that EBC volume is strongly correlated to total expired air volume, and that EBC volume does not correlate with measures of airflow obstruction. The percentage of total water vapor removed by the condensation process was calculated to be approximately 40% using the Jaeger collection system. They reported protein concentrations of approximately 12 $\mu\text{g}/\text{ml}$ of EBC both in subjects with COPD and control subjects. There was tighter correlation of protein with volume in control subjects than in patients

with COPD. Having extensively studied EBC during mechanical ventilation, they reported normal or increased concentrations of protein and urea, despite larger volumes of EBC collected from intubated subjects than from subjects spontaneously breathing. This suggests either that the additional condensate volume collected while intubated is paralleled by increased ALF trapping, or that protein levels in ALF increase during mechanical ventilation.

Dr. Corradi has shown that EBC malondialdehyde and GSH concentrations are essentially independent of expiratory flow rate in the range of 50 to 200 ml s^{-1} . Dr. Hunt has shown that the pH of EBC is not affected by ventilation pattern (hyperventilation/hypoventilation), methacholine-induced airflow limitation, temperature or duration of collection, condenser material, or duration of storage (170). Hydrogen peroxide levels have been reported to be dependent on exhaled flow rates (176) in a manner reflective of exhaled NO (lower flow, higher concentrations) (72).

It may be prudent, and of use in the near future, to begin the standardization process by following some of Dr. Becher's suggestions. Specifically, he recommends presampling inspection of mouth, teeth, tongue, and throat to identify inflammation and spirometry to correlate with EBC assay results, unless it has been shown that expiratory flow issues are not relevant to the substance of interest. It may be useful to rinse the mouth (e.g., two times with 30 ml water) before collection, although again this needs to be studied for each marker of interest.

Although the use of nose clips for F_{ENO} measurements has been discouraged because they can enhance nasal contamination of the orally exhaled airstream with NO, this logic does not extend directly to the process of EBC collection. Samples of EBC are collected over many minutes, involving multiple, full respiratory cycles. This contrasts with the single exhalation-only technique usually used for F_{ENO} measures. For EBC collections, nose clips might increase entrained NO during exhalation, while decreasing it during inhalation. The data are lacking to support an unequivocal recommendation regarding nose clips during EBC collection, but as this issue is considered, it will be necessary to note that extension of F_{ENO} logic to EBC sampling is not straightforward.

Other early recommendations include encouragement for the subject to breathe at normal tidal volumes and avoidance of hyperventilation, and removal of collector from mouth during coughing or burping. There may be advantages to constant monitoring of airflow with biofeedback during collection, as it may assist the subject in performing the collection maneuver consistently. Such online registration of breathing frequency, tidal volume, mean expiratory flow, and minute volume might be helpful for comparing EBC constituents over time in one patient. It was suggested that forced lung function maneuvers should not be performed within 10 min before the collection. There is no consensus in these recommendations. Data to support any mandates in this regard are lacking. We stress the importance of examining the potential confounders for each individual substance of interest, and encourage continued empiric investigation to determine which potential confounders indeed are sufficiently problematic to warrant adding complexity to the collection technique.

The optimum temperature in which to store EBC samples is an issue that serves to underscore the diverse characteristics of EBC constituents. Nitrate is considered a stable compound in the matrix of EBC, and therefore will not react away at any reasonable temperature. However, NO_2^- has the potential to be oxidized to NO_3^- or, for that matter, reduced to NO over time in the chemical matrix of EBC. The reduction of NO_2^- occurs more aggressively at low pH, and therefore the NO_2^- stability is pH dependent. Hydrogen peroxide will react with

lipids and inorganic molecules, and can be expected to decline over time. However, data examining storage at -70°C suggest that this may be acceptable for even up to 40 days (179). The pH of EBC in storage at -20° did not change significantly over periods longer than 1 year (165). For the less stable compounds, it seems wise to store the samples in as cold a temperature as possible and assay soon after collection.

To assist with interlaboratory communications, it will be helpful if reports of EBC assays include as many methodologic details as reasonably possible regarding the type of condenser, temperature and duration of collections, and any other breathing patterns used other than normal tidal respiration.

An ERS/ATS joint task force on EBC has begun to address these issues. One key advantage of EBC assays is the simplicity of the collection process. Although many factors may affect concentrations of substances in EBC, empiric investigations into these factors need to be performed. For now, the dependence on collection technique of each disease marker studied needs to be evaluated by the investigators involved.

SECTION SUMMARY: EBC, REDOX STATUS, AND NITROGEN OXIDES

Still clouded by uncertainties regarding anatomic sources and complex chemistries, EBC nonetheless allows us to gain cautious insights into ALF constituents in disease states and in health. Lung redox information has become obtainable through this technology, through assays of pH, nitrogen oxides, hydrogen peroxide, glutathione, and markers of the presence of disturbances in these compounds, such as nitrotyrosine, isoprostane, and aldehydes. EBC has the potential to complement Fe_{NO} to more fully elucidate the nitrogen oxide chemistry of the airways and lung. As a marker of disease activity, EBC assays may monitor acid stress, oxidative stress, and inflammation, separate but often-related entities to which many of our therapies are directed.

OVERALL SUMMARY OF WORKSHOP

This meeting was convened to encourage cooperation among international investigators interested in assessment of airway nitrogen oxides and redox chemistry. Methodology of exhaled NO measurements was extensively discussed. Updates to NO sensor technologies were presented, the ATS current online method for NO measurement was reviewed in light of recent experience, and changes were proposed that are currently being incorporated in a revision to the ATS statement. The role of nasal NO measurement was discussed and the new fields of NO excretion modeling and NO measurement in ventilated patients were presented. In addition, the less technically defined methodology of EBC collection and assay were discussed, stressing primarily redox relevant chemistries. Although assays of large molecules in EBC, such as cytokines, suffer from insufficiently sensitive assays, the sensitivity of the assays available for the redox-relevant compounds of central interest to this workshop are clearly sufficient for the purpose. Issues of dilution remain critically important for an understanding of precise chemical constituents of the airway, although ratios of compounds nonetheless can provide insights into overall lung redox status. Potential for contamination from the upper airway remains a concern in all oral exhaled biomarker measurements. Nonetheless, the data supporting the clinical utility of Fe_{NO} measures now are substantial. Although EBC studies are being rapidly published, there are numerous biomarkers present, and thus technical validation remains less advanced than for Fe_{NO} . It is reasonable to conclude that, as the technical work is performed, concurrent

measures of Fe_{NO} , EBC nitrogen oxides, pH, hydrogen peroxide, and isoprostanes and other markers of redox activity in the lung will enhance our understanding of nitrogen oxide balance and the pathologic process we call "oxidative stress."

THIS OFFICIAL ATS WORKSHOP PROCEEDINGS WAS DEVELOPED BY AN AD HOC SUBCOMMITTEE OF THE ASSEMBLY ON ALLERGY, IMMUNOLOGY, AND INFLAMMATION.

Writing Committee

Background: PHILIP E. SILKOFF, M.D.
 Sensor technologies: SERPIL C. ERZURUM, M.D.
 Online exhaled NO measurement: PHILIP E. SILKOFF, M.D.
 Clinical application of exhaled NO: PHILIP E. SILKOFF, M.D.
 Nasal NO: JON O. LUNDBERG, Ph.D.
 Modeling of NO excretion: STEVEN C. GEORGE, M.D., Ph.D.
 NO measurement in ventilated patients: NANDOR MARCZIN, M.D., Ph.D.
 Exhaled breath condensate: JOHN F. HUNT, M.D.; RICHARD EFFROS, M.D.; ILDIKO HORVATH, M.D.; SERPIL C. ERZURUM, M.D.

Conflict of Interest Statement: R.M.E. received funds (\$100,000) from Pfizer to develop methods for interpreting exhaled breath condensate data in 2003 and \$20,000 for consultation with Pfizer in 2005 on issues involving condensates. S.C.E. does not have a financial relationship with a commercial entity that has an interest in the subject of this manuscript. S.C.G. has received a NIOX instrument as a gift from Aerocrine AB, and has patents issued and pending related to exhaled NO, for which Aerocrine AB has entered into a licensing agreement with the University of California, Irvine; he has received a total of \$1,300 from this license agreement. I.H. does not have a financial relationship with a commercial entity that has an interest in the subject of this manuscript. J.H. is a cofounder and substantial shareholder of Respiratory Research, Inc, which designs and manufactures exhaled breath condensate collection equipment and has licensed exhaled breath condensate pH and other assays from the University of Virginia; he is an inventor of exhaled breath condensate nitrogen oxide and pH assays. J.L. is a shareholder in Aerocrine AB Sweden, a company that manufactures and sells equipment for exhaled NO measurements. N.M. does not have a financial relationship with a commercial entity that has an interest in the subject of this manuscript. P.E.S. is currently a full-time employee of AstraZeneca, but this bears no relationship or conflict of interest to this document; previously, he was a paid consultant for Ionics-Sievers Instruments and Aerocrine AB, manufacturers of exhaled NO meters; he receives royalties from Ionics-Sievers Instruments and Apero Biosystems for licensed patents.

Workshop Participants

PHILIP E. SILKOFF, M.D., United States (Chair)
 SERPIL C. ERZURUM, M.D., United States (Chair)
 JOHN F. HUNT, M.D., United States (Chair)
 KJELL H. ALVING, Ph.D., Sweden
 EUGENIO BARALDI, M.D., Italy
 JOSE M. CHATKIN, M.D., Brazil
 MASSIMO CORRADI, M.D., Italy
 AARON DEYKIN, M.D., United States
 RAED A. DWEIK, M.D., United States
 RICHARD EFFROS, M.D., United States
 W. MICHAEL FOSTER, Ph.D., United States
 BEN GASTON, M.D., United States
 STEVEN C. GEORGE, M.D., Ph.D., United States
 CHRISTIAN GESSNER, M.D., Germany
 GIANFRANCO GIUBILEO, M.D., Italy
 MICHAEL GOLDMAN, M.D., United States
 CARLOS GUTIERREZ, M.D., Canada
 MARIEANN HOGMAN, Ph.D., Sweden
 JENS HOHLFELD, M.D., Germany
 OLAF HOLZ, Germany
 ILDIKO HORVATH, M.D., Hungary
 DANIEL W. LASKOWSKI, R.P.F.T., United States
 JON O. LUNDBERG, Ph.D., Sweden
 NANDOR MARCZIN, M.D., Ph.D., United Kingdom
 KAREN McRAE, M.D., Canada
 SANJAY MEHTA, M.D., Canada
 ANNA-CARIN OLIN, M.D., Sweden
 SOLBERT PERMUTT, M.D., United States
 ANTHONY REDINGTON, M.D., United Kingdom
 TERENCE RISBY, Ph.D., United States
 RICHARD A. ROBBINS, M.D., United States
 JIGME SETHI, M.D., United States
 ROBERT S. TEPPER, M.D., Ph.D., United States
 EDDIE WEITZBERG, M.D., Sweden
 HUBERT WIRTZ, M.D., Germany

Company Representatives: Aerocrine AB, Sweden: MATS CARLSON; Aerocrine, Inc., United States: TREVOR BOURKE, IAN McLEOD; Eco Physics, United States: TOM McKARNS; Eco Medics, Switzerland: DIRK WENDT; EKIPS Technologies, United States: PATRICK McCANN; FILT Lungen- und Thoraxdiagnostik, Germany: GUNTHER

BECHER; Ionics Instruments, United States: RIC HUTTE; Jaeger, Germany: WERNER R. STEINHAEUSSER, GUNTHER BECHER; Methapharm, Canada: KAREN MICULA

References

- American Thoracic Society. Recommendations for standardized procedures for the on-line and off-line measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide in adults and children—1999. *Am J Respir Crit Care Med* 1999;160:2104–2117.
- American Thoracic Society/European Respiratory Society. ATS/ERS recommendations for standardized procedures for the online and offline measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide, 2005. *Am J Respir Crit Care Med* 2005;171:912–930.
- Horvath I, Hunt J, Barnes PJ. Exhaled breath condensate: methodological recommendations and unresolved questions. *Eur Respir J* 2005;26:523–548.
- Bates CA, Silkoff PE. Exhaled nitric oxide in asthma: from bench to bedside. *J Allergy Clin Immunol* 2003;111:256–262.
- Baraldi E, Scollo M, Zaramella C, Zanconato S, Zacchello F. A simple flow-driven method for online measurement of exhaled NO starting at the age of 4 to 5 years. *Am J Respir Crit Care Med* 2000;162:1828–1832.
- Deykin A, Massaro AF, Drazen JM, Israel E. Exhaled nitric oxide as a diagnostic test for asthma: online versus offline techniques and effect of flow rate. *Am J Respir Crit Care Med* 2002;165:1597–1601.
- Gilain L, Bedu M, Jouaville L, Guichard C, Advenier D, Mom T, Laurent S, Caillaud D. Analysis of nasal and exhaled nitric oxide concentration in nasal polyposis [in French]. *Ann Otolaryngol Chir Cervicofac* 2002;119:234–242.
- Girgis RE, Qureshi MA, Abrams J, Swerdlow P. Decreased exhaled nitric oxide in sickle cell disease: relationship with chronic lung involvement. *Am J Hematol* 2003;72:177–184.
- Ho LP, Innes JA, Greening AP. Nitrite levels in breath condensate of patients with cystic fibrosis is elevated in contrast to exhaled nitric oxide. *Thorax* 1998;53:680–684.
- Jobsis Q, Raatgeep HC, Hop WC, de Jongste JC. Controlled low flow off line sampling of exhaled nitric oxide in children. *Thorax* 2001;56:285–289.
- Kissoon N, Duckworth LJ, Blake KV, Murphy SP, Taylor CL, Silkoff PE. Fe(NO): relationship to exhalation rates and online versus bag collection in healthy adolescents. *Am J Respir Crit Care Med* 2000;162:539–545.
- Kissoon N, Duckworth LJ, Blake KV, Murphy SP, Lima JJ. Effect of beta2-agonist treatment and spirometry on exhaled nitric oxide in healthy children and children with asthma. *Pediatr Pulmonol* 2002;34:203–208.
- Kissoon N, Duckworth LJ, Blake KV, Murphy SP, Taylor CL, DeNicola LR, Silkoff PE. Exhaled nitric oxide concentrations: online versus offline values in healthy children. *Pediatr Pulmonol* 2002;33:283–292.
- Pedroletti C, Zetterquist W, Nordvall L, Alving K. Evaluation of exhaled nitric oxide in schoolchildren at different exhalation flow rates. *Pediatr Res* 2002;52:393–398.
- Shin HW, Rose-Gottron CM, Sufi RS, Perez F, Cooper DM, Wilson AF, George SC. Flow-independent nitric oxide exchange parameters in cystic fibrosis. *Am J Respir Crit Care Med* 2002;165:349–357.
- Silkoff PE, McClean PA, Slutsky AS, Furlott HG, Hoffstein E, Wakita S, Chapman KR, Szalai JP, Zamel N. Marked flow-dependence of exhaled nitric oxide using a new technique to exclude nasal nitric oxide. *Am J Respir Crit Care Med* 1997;155:260–267.
- St. Croix CM, Wetter TJ, Pegelow DF, Meyer KC, Dempsey JA. Assessment of nitric oxide formation during exercise. *Am J Respir Crit Care Med* 1999;159:1125–1133.
- van der Lee I, van den Bosch JM, Zanen P. Reduction of variability of exhaled nitric oxide in healthy volunteers. *Respir Med* 2002;96:1014–1020.
- Wildhaber JH, Hall GL, Stick SM. Measurements of exhaled nitric oxide with the single-breath technique and positive expiratory pressure in infants. *Am J Respir Crit Care Med* 1999;159:74–78.
- Kharitonov S, Alving K, Barnes PJ. Exhaled and nasal nitric oxide measurements: recommendations. The European Respiratory Society Task Force. *Eur Respir J* 1997;10:1683–1693.
- Kharitonov SA, Gonio F, Kelly C, Meah S, Barnes PJ. Reproducibility of exhaled nitric oxide measurements in healthy and asthmatic adults and children. *Eur Respir J* 2003;21:433–438.
- Persson MG, Lonnqvist PA, Gustafsson LE. Positive end-expiratory pressure ventilation elicits increases in endogenously formed nitric oxide as detected in air exhaled by rabbits. *Anesthesiology* 1995;82:969–974.
- Kondo R, Haniuda M, Yamanda T, Sato E, Fujimoto K, Kubo K, Amano J. Effects of expiratory pressure on nitric oxide in exhaled breath: is exhaled nitric oxide really unaffected by pressure? *Respir Physiol Neurobiol* 2003;139:33–40.
- Alving K, Weitzberg E, Lundberg JM. Increased amount of nitric oxide in exhaled air of asthmatics. *Eur Respir J* 1993;6:1368–1370.
- Kharitonov SA, Yates D, Springall DR, Buttery L, Polak J, Robbins RA, Barnes PJ. Exhaled nitric oxide is increased in asthma. *Chest* 1995;107:156S–157S.
- Yates DH, Kharitonov SA, Thomas PS, Barnes PJ. Endogenous nitric oxide is decreased in asthmatic patients by an inhibitor of inducible nitric oxide synthase. *Am J Respir Crit Care Med* 1996;154:247–250.
- Baraldi E, Azzolin NM, Zanconato S, Dario C, Zacchello F. Corticosteroids decrease exhaled nitric oxide in children with acute asthma. *J Pediatr* 1997;131:381–385.
- Silkoff PE, McClean P, Spino M, Erlich L, Slutsky AS, Zamel N. Dose-response relationship and reproducibility of the fall in exhaled nitric oxide after inhaled beclomethasone dipropionate therapy in asthma patients. *Chest* 2001;119:1322–1328.
- Silkoff PE, McClean PA, Slutsky AS, Caramori M, Chapman KR, Gutierrez C, Zamel N. Exhaled nitric oxide and bronchial reactivity during and after inhaled beclomethasone in mild asthma. *J Asthma* 1998;35:473–479.
- Beck-Ripp J, Griese M, Arenz S, Koring C, Pasqualoni B, Bifulco P. Changes of exhaled nitric oxide during steroid treatment of childhood asthma. *Eur Respir J* 2002;19:1015–1019.
- Buchvald F, Eiberg H, Bisgaard H. Heterogeneity of FeNO response to inhaled steroid in asthmatic children. *Clin Exp Allergy* 2003;33:1735–1740.
- Kharitonov SA, Yates DH, Barnes PJ. Inhaled glucocorticoids decrease nitric oxide in exhaled air of asthmatic patients. *Am J Respir Crit Care Med* 1996;153:454–457.
- Bisgaard H, Loland L, Oj JA. NO in exhaled air of asthmatic children is reduced by the leukotriene receptor antagonist montelukast. *Am J Respir Crit Care Med* 1999;160:1227–1231.
- Silkoff PE, Romero FA, Gupta N, Townley RG, Milgrom H. Exhaled nitric oxide in children with asthma receiving Xolair (omalizumab), a monoclonal anti-immunoglobulin E antibody. *Pediatrics* 2004;113:e308–e312.
- Buchvald F, Bisgaard H. Comparisons of the complementary effect on exhaled nitric oxide of salmeterol vs montelukast in asthmatic children taking regular inhaled budesonide. *Ann Allergy Asthma Immunol* 2003;91:309–313.
- Warke TJ, Fitch PS, Brown V, Taylor R, Lyons JD, Ennis M, Shields MD. Exhaled nitric oxide correlates with airway eosinophils in childhood asthma. *Thorax* 2002;57:383–387.
- van den Toorn LM, Overbeek SE, de Jongste JC, Leman K, Hoogsteden HC, Prins JB. Airway inflammation is present during clinical remission of atopic asthma. *Am J Respir Crit Care Med* 2001;164:2107–2113.
- Jatakanon A, Lim S, Barnes PJ. Changes in sputum eosinophils predict loss of asthma control. *Am J Respir Crit Care Med* 2000;161:64–72.
- Piacentini GL, Bodini A, Costella S, Vicentini L, Mazzi P, Sperandio S, Boner AL. Exhaled nitric oxide and sputum eosinophil markers of inflammation in asthmatic children. *Eur Respir J* 1999;13:1386–1390.
- Mahut B, Delclaux C, Tillie-Leblond I, Gosset P, Delacourt C, Zerah-Lancner F, Harf A, de Blic J. Both inflammation and remodeling influence nitric oxide output in children with refractory asthma. *J Allergy Clin Immunol* 2004;113:252–256.
- Leuppi JD, Salome CM, Jenkins CR, Anderson SD, Xuan W, Marks GB, Koskela H, Brannan JD, Freed R, Andersson M, et al. Predictive markers of asthma exacerbation during stepwise dose reduction of inhaled corticosteroids. *Am J Respir Crit Care Med* 2001;163:406–412.
- Lanz MJ, Leung DY, McCormick DR, Harbeck R, Szefer SJ, White CW. Comparison of exhaled nitric oxide, serum eosinophilic cationic protein, and soluble interleukin-2 receptor in exacerbations of pediatric asthma. *Pediatr Pulmonol* 1997;24:305–311.
- Harkins MS, Fiato KL, Iwamoto GK. Exhaled nitric oxide predicts asthma exacerbation. *J Asthma* 2004;41:471–476.
- Jones SL, Kittelson J, Cowan JO, Flannery EM, Hancox RJ, McLachlan CR, Taylor DR. The predictive value of exhaled nitric oxide measurements in assessing changes in asthma control. *Am J Respir Crit Care Med* 2001;164:738–743.
- Karadag B, James AJ, Gultekin E, Wilson NM, Bush A. Nasal and lower airway level of nitric oxide in children with primary ciliary dyskinesia. *Eur Respir J* 1999;13:1402–1405.
- Alving K, Weitzberg E, Lundberg JM. Increased amount of nitric oxide in exhaled air of asthmatics. *Eur Respir J* 1993;6:1368–1370.

47. Lundberg JO, Weitzberg E, Nordvall SL, Kuylenstierna R, Lundberg JM, Alving K. Primarily nasal origin of exhaled nitric oxide and absence in Kartagener's syndrome. *Eur Respir J* 1994;7:1501-1504.
48. Schedin U, Frostell C, Persson MG, Jakobsson J, Andersson G, Gustafsson LE. Contribution from upper and lower airways to exhaled endogenous nitric oxide in humans. *Acta Anaesthesiol Scand* 1995;39:327-332.
49. Lundberg JON, Farkas-Szallasi T, Weitzberg E, Rinder J, Lidholm J, Ånggård A, Hökfelt T, Lundberg JM, Alving K. High nitric oxide production in human paranasal sinuses. *Nat Med* 1995;1:370-373.
50. Runer T, Cervin A, Lindberg S, Uddman R. Nitric oxide is a regulator of muco ciliary activity in the upper respiratory tract. *Otolaryngol Head Neck Surg* 1998;107:40-46.
51. Lundberg JO, Nordvall SL, Weitzberg E, Kollberg H, Alving K. Exhaled nitric oxide in paediatric asthma and cystic fibrosis. *Arch Dis Child* 1996;75:323-326.
52. Balfour-Lynn I, Laverty A, Dinwiddle R. Reduced upper airway nitric oxide in cystic fibrosis. *Arch Dis Child* 1996;75:319-322.
53. Kharitonov S, Rajakulasingam K, O'Connor B, Durham S, Barnes P. Nasal nitric oxide is increased in patients with asthma and allergic rhinitis and may be modulated by nasal glucocorticosteroids. *J Allergy Clin Immunol* 1997;99:58-64.
54. Arnal JF, Didier A, Rami I, M'Rini C, Charlet JP, Serrano E, Besombes JP. Nasal nitric oxide is increased in allergic rhinitis. *Clin Exp Allergy* 1997;27(4):358-362.
55. Martin U, Bryden K, Devoy M, Howarth P. Increased levels of exhaled nitric oxide during nasal and oral breathing in subjects with seasonal rhinitis. *J Allergy Clin Immunol* 1996;97:768-772.
56. Lundberg JO, Weitzberg E. Nasal nitric oxide in man. *Thorax* 1999;54:947-952.
57. Djupesland PG, Chatkin JM, Qian W, Haight JS. Nitric oxide in the nasal airway: a new dimension in otorhinolaryngology. *Am J Otolaryngol* 2001;22:19-32.
58. Lundberg JO, Weitzberg E, Lundberg JM, Alving K. Nitric oxide in exhaled air. *Eur Respir J* 1996;9:2671-2680.
59. Narang I, Ersu R, Wilson NM, Bush A. Nitric oxide in chronic airway inflammation in children: diagnostic use and pathophysiological significance. *Thorax* 2002;57:586-589.
60. Wodehouse T, Kharitonov SA, Mackay IS, Barnes PJ, Wilson R, Cole PJ. Nasal nitric oxide measurements for the screening of primary ciliary dyskinesia. *Eur Respir J* 2003;21:43-47.
61. Horvath I, Loukides S, Wodehouse T, Csiszer E, Cole PJ, Kharitonov SA, Barnes PJ. Comparison of exhaled and nasal nitric oxide and exhaled carbon monoxide levels in bronchiectatic patients with and without primary ciliary dyskinesia. *Thorax* 2003;58:68-72.
62. Coren ME, Meeks M, Morrison I, Buchdahl RM, Bush A. Primary ciliary dyskinesia: age at diagnosis and symptom history. *Acta Paediatr* 2002;91:667-669.
63. Bush A, Cole P, Hariri M, Mackay I, Phillips G, O'Callaghan C, Wilson R, Warner JO. Primary ciliary dyskinesia: diagnosis and standards of care. *Eur Respir J* 1998;12:982-988.
64. Bush A. Primary ciliary dyskinesia. *Acta Otorhinolaryngol Belg* 2000;54:317-324.
65. Silkoff PE, Robbins RA, Gaston B, Lundberg JO, Townley RG. Endogenous nitric oxide in allergic airway disease. *J Allergy Clin Immunol* 2000;105:438-448.
66. Palm JP, Graf P, Lundberg JO, Alving K. Characterization of exhaled nitric oxide: introducing a new reproducible method for nasal nitric oxide measurements. *Eur Respir J* 2000;16:236-241.
67. Maniscalco M, Weitzberg E, Sundberg J, Sofia M, Lundberg JO. Assessment of nasal and sinus nitric oxide output using single-breath humming exhalations. *Eur Respir J* 2003;22:323-329.
68. Weitzberg E, Lundberg JO. Humming greatly increases nasal nitric oxide. *Am J Respir Crit Care Med* 2002;166:144-145.
69. Lundberg JO, Maniscalco M, Sofia M, Lundblad L, Weitzberg E, Maniscalco M. Humming, nitric oxide, and paranasal sinus obstruction. *JAMA* 2003;289:302-303.
70. Gustafsson LE, Leone AM, Persson MG, Wiklund NP, Moncada S. Endogenous nitric oxide is present in the exhaled air of rabbits, guinea pigs and humans. *Biochem Biophys Res Commun* 1991;181:852-857.
71. Hogman M, Stromberg S, Schedin U, Frostell C, Hedenstierna G, Gustafsson LE. Nitric oxide from the human respiratory tract efficiently quantified by standardised single breath measurements. *Acta Physiol Scand* 1997;159:345-346.
72. Silkoff PE, McClean PA, Slutsky AS, Furlott HG, Hoffstein E, Wakita S, Chapman KR, Szalai JP, Zamel N. Marked flow-dependence of exhaled nitric oxide using a new technique to exclude nasal nitric oxide. *Am J Respir Crit Care Med* 1997;155:260-267.
73. Tsoukias NM, George SC. A two-compartment model of pulmonary nitric oxide exchange dynamics. *J Appl Physiol* 1998;85:653-666.
74. Hogman M, Draic N, Ehrstedt C, Merilainen P. Exhaled nitric oxide partitioned into alveolar, lower airways and nasal contributions. *Respir Med* 2000;94:985-991.
75. Pietropaoli AP, Perillo IB, Torres A, Perkins PT, Frasier LM, Utell MJ, Frampton MW, Hyde RW. Simultaneous measurement of nitric oxide production by conducting and alveolar airways of humans. *J Appl Physiol* 1999;87:1532-1542.
76. Silkoff PE, Sylvester JT, Zamel N, Permutt S. Airway nitric oxide diffusion in asthma: role in pulmonary function and bronchial responsiveness. *Am J Respir Crit Care Med* 2000;161:1218-1228.
77. Hyde RW, Geigel EJ, Olszowka AJ, Krasney JA, Forster RE II, Utell MJ, Frampton MW. Determination of production of nitric oxide by lower airways of humans: theory. *J Appl Physiol* 1997;82:1290-1296.
78. Tsoukias NM, Dabdub D, Wilson AF, George SC. Effect of alveolar volume and sequential filling on the diffusing capacity of the lungs: I. theory. *Respir Physiol* 2000;120:231-250.
79. Tsoukias NM, Wilson AF, George SC. Effect of alveolar volume and sequential filling on the diffusing capacity of the lungs: II. experiment. *Respir Physiol* 2000;120:251-271.
80. Guenard H, Varenne N, Vaida P. Determination of lung capillary blood volume and membrane diffusing capacity in man by the measurements of NO and CO transfer. *Respir Physiol* 1987;70:113-120.
81. Borland CDR, Higenbottam TW. A simultaneous single breath measurement of pulmonary diffusing capacity with nitric oxide and carbon monoxide. *Eur Respir J* 1989;2:56-63.
82. Tsoukias NM, Shin H-W, Wilson AF, George SC. A single breath technique with variable flow rate to characterize nitric oxide exchange dynamics in the lungs. *J Appl Physiol* 2001;91:477-487.
83. Tsoukias NM, Tannous Z, Wilson AF, George SC. Single-exhalation profiles of NO and CO₂ in humans: effect of dynamically changing flow rate. *J Appl Physiol* 1998;85:642-652.
84. Condorelli P, Shin HW, George SC. Characterizing airway and alveolar nitric oxide exchange during tidal breathing using a three-compartment model. *J Appl Physiol* 2004;96:1832-1842.
85. Shin HW, George SC. Impact of axial diffusion on nitric oxide exchange in the lungs. *J Appl Physiol* 2002;93:2070-2080.
86. Shin HW, Condorelli P, Rose-Gottron CM, Cooper DM, George SC. Probing the impact of axial diffusion on nitric oxide exchange dynamics with heliox. *J Appl Physiol* 2004;97:874-882.
87. Van Muylem A, Noel C, Paiva M. Modeling of impact of gas molecular diffusion on nitric oxide expired profile. *J Appl Physiol* 2002.
88. Hogman M, Holmkvist T, Wegener T, Emtner M, Andersson M, Hedenstrom H, Merilainen P. Extended NO analysis applied to patients with COPD, allergic asthma and allergic rhinitis. *Respir Med* 2002;96:24-30.
89. Lehtimäki L, Turjanmaa V, Kankaanranta H, Saarelainen S, Hahtola P, Moilanen E. Increased bronchial nitric oxide production in patients with asthma measured with a novel method of different exhalation flow rates. *Ann Med* 2000;32:417-423.
90. Lehtimäki L, Kankaanranta H, Saarelainen S, Turjanmaa V, Moilanen E. Inhaled fluticasone decreases bronchial but not alveolar nitric oxide output in asthma. *Eur Respir J* 2001;18:635-639.
91. Lehtimäki L, Kankaanranta H, Saarelainen S, Hahtola P, Järvenpää R, Koivula T, Turjanmaa V, Moilanen E. Extended exhaled NO measurement differentiates between alveolar and bronchial inflammation. *Am J Respir Crit Care Med* 2001;163:1557-1561.
92. Shin HW, Rose-Gottron CM, Cooper DM, Hill M, George SC. Impact of high-intensity exercise on nitric oxide exchange in healthy adults. *Med Sci Sports Exerc* 2003;35:995-1003.
93. Shin H-W, Rose-Gottron CM, Cooper DM, Newcombe RL, George SC. Airway diffusing capacity of nitric oxide is not impacted by steroid therapy in asthma. *J Appl Physiol* 2004;96:65-75.
94. Girgis RE, Gughani MK, Abrams J, Mayes MD. Partitioning of alveolar and conducting airway nitric oxide in scleroderma lung disease. *Am J Respir Crit Care Med* 2002;165:1587-1591.
95. Hogman M, Holmkvist T, Walinder R, Merilainen P, Ludviksdottir D, Hakansson L, Hedenstrom H. Increased nitric oxide elimination from the airways after smoking cessation. *Clin Sci (Lond)* 2002;103:15-19.
96. Borner U, Klimek M. Does an analysis of exhaled air indicate the metabolic state of critically ill patients? *Intensive Care Med* 1998;24(5):403-404.

97. Scholpp J, Schubert JK, Miekisch W, Geiger K. Breath markers and soluble lipid peroxidation markers in critically ill patients. *Clin Chem Lab Med* 2002;40(6):587-594.
98. Carpenter CT, Price PV, Christman BW. Exhaled breath condensate isoprostanes are elevated in patients with acute lung injury or ARDS. *Chest* 1998;114(6):1653-1659.
99. Wilson WC, Swetland JF, Benumof JL, Laborde P, Taylor R. General anesthesia and exhaled breath hydrogen peroxide. *Anesthesiology* 1992;76(5):703-710.
100. Zegdi R, Perrin D, Burdin M, Boiteau R, Tenaillon A. Increased endogenous carbon monoxide production in severe sepsis. *Intensive Care Med* 2002;28(6):793-796.
101. Andreoni KA, Kazui M, Cameron DE, Nyhan D, Sehnert SS, Rohde CA, Bulkley GB, Risby TH. Ethane: a marker of lipid peroxidation during cardiopulmonary bypass in humans. *Free Radic Biol Med* 1999;26(3-4):439-445.
102. Kazui M, Andreoni KA, Williams GM, Perler BA, Bulkley GB, Beattie C, Donham RT, Sehnert SS, Burdick JF, Risby TH. Visceral lipid peroxidation occurs at reperfusion after supraceliac aortic cross-clamping. *J Vasc Surgery* 1994;19(3):473-477.
103. Ishibe Y, Liu R, Hirotsawa J, Kawamura K, Yamasaki K, Saito N. Exhaled nitric oxide level decreases after cardiopulmonary bypass in adult patients. *Crit Care Med* 2000;28(12):3823-3827.
104. Marczin N, Kovesi T, Royston D. Exhaled nitric oxide as a marker of lung injury in coronary artery bypass surgery. *Br J Anaesthesia* 2003;90(1):101-104.
105. Adding LC, Bannenberg GL, Gustafsson LE. Gadolinium chloride inhibition of pulmonary nitric oxide production and effects on pulmonary circulation in the rabbit. *Pharmacol Toxicol* 1998;83(1):8-15.
106. Dweik RA, Laskowski D, Abu-Soud HM, Kaneko F, Hutte R, Stuehr DJ, Erzurum SC. Nitric oxide synthesis in the lung: regulation by oxygen through a kinetic mechanism. *J Clin Invest* 1998;101(3):660-666.
107. Marczin N, Bundy RE, Hoare GS, Yacoub M. Redox regulation following cardiac ischemia and reperfusion. *Coron Artery Dis* 2003;14(2):123-133.
108. Marczin N, Riedel B, Royston D, Yacoub M. Intravenous nitrate vasodilators and exhaled nitric oxide. *Lancet* 1997;349(9067):1742.
109. Malmstrom RE, Tornberg D, Settergren G, Liska J, Angdin M, Lundberg JO, Weitzberg E. Endogenous nitric oxide release by vasoactive drugs monitored in exhaled air. *Am J Respir Crit Care Med* 2003;168:114-120.
110. Cremona G, Higenbottam T, Takao M, Hall L, Bower EA. Exhaled nitric oxide in isolated pig lungs. *J Appl Physiology* 1995;78(1):59-63.
111. Persson MG, Agvald P, Gustafsson LE. Detection of nitric oxide in exhaled air during administration of nitroglycerin in vivo. *Br J Pharmacol* 1994;111(3):825-828.
112. Husain M, Adrie C, Ichinose F, Kavosi M, Zapol WM. Exhaled nitric oxide as a marker for organic nitrate tolerance. *Circulation* 1994;89(6):2498-2502.
113. Busch T, Bartsch P, Pappert D, Grunig E, Hildebrandt W, Elser H, Falke KJ, Swenson ER. Hypoxia decreases exhaled nitric oxide in mountaineers susceptible to high-altitude pulmonary edema. *Am J Respir Crit Care Med* 2001;163(2):368-373.
114. Cucchiari G, Tatum AH, Brown MC, Camporesi EM, Daucher JW, Hakim TS. Inducible nitric oxide synthase in the lung and exhaled nitric oxide after hyperoxia. *Am J Physiol* 1999;277(3 Pt 1):L636-L644.
115. Schmetterer L, Strenn K, Kastner J, Eichler HG, Wolzt M. Exhaled NO during graded changes in inhaled oxygen in man. *Thorax* 1997;52(8):736-738.
116. Tsuchiya M, Tokai H, Takehara Y, Haraguchi Y, Asada A, Utsumi K, Inoue M. Interrelation between oxygen tension and nitric oxide in the respiratory system. *Am J Respir Crit Care Med* 2000;162(4 Pt 1):1257-1261.
117. Hall GL, Reinmann B, Wildhaber JH, Frey U. Tidal exhaled nitric oxide in healthy, unsedated newborn infants with prenatal tobacco exposure. *J Appl Physiol* 2002;92(1):59-66.
118. Persson MG, Lonnqvist PA, Gustafsson LE. Positive end-expiratory pressure ventilation elicits increases in endogenously formed nitric oxide as detected in air exhaled by rabbits. *Anesthesiology* 1995;82(4):969-974.
119. Carlin RE, Ferrario L, Boyd JT, Camporesi EM, McGraw DJ, Hakim TS. Determinants of nitric oxide in exhaled gas in the isolated rabbit lung. *Am J Respir Crit Care Med* 1997;155(3):922-927.
120. Marczin N, Jilling T, Papapetropoulos A, Go C, Catravas JD. Cytoskeleton-dependent activation of the inducible nitric oxide synthase in cultured aortic smooth muscle cells. *Br J Pharmacol* 1996;118(5):1085-1094.
121. Forsberg S, Ludwigs U, Hedenstierna G. Effects of ventilatory pattern on exhaled nitric oxide in mechanically ventilated rabbits. *Acta Anaesthesiol Scand* 1999;43(4):464-469.
122. Artlich A, Adding C, Agvald P, Persson MG, Lonnqvist PA, Gustafsson LE. Exhaled nitric oxide increases during high frequency oscillatory ventilation in rabbits. *Exp Physiol* 1999;84(5):959-969.
123. Carlin RE, McGraw DJ, Camporesi EM, Hakim TS. Increased nitric oxide in exhaled gas is an early marker of hypovolemic states. *J Surg Res* 1997;69(2):362-366.
124. Fernandez-Mondejar E, Hambræus-Jonzon K, Roneus A, Hedenstierna G. Nitric oxide increases dramatically in air exhaled from lung regions with occluded vessels. *Acta Anaesthesiol Scand* 2003;47(3):312-318.
125. Beghetti M, Silkoff PE, Caramori M, Holtby HM, Slutsky AS, Adatia I. Decreased exhaled nitric oxide may be a marker of cardiopulmonary bypass-induced injury. *Ann Thoracic Surg* 1998;66(2):532-534.
126. Tworetzky W, Moore P, Bekker JM, Bristow J, Black SM, Fineman JR. Pulmonary blood flow alters nitric oxide production in patients undergoing device closure of atrial septal defects. *J Am Coll Cardiol* 2000;35(2):463-467.
127. Massaro AF, Drazen JM. Exhaled nitric oxide during exercise: site of release and modulation by ventilation and blood flow. *J Appl Physiol* 1996;80(6):1863-1864.
128. Tornberg DC, Bjorne H, Lundberg JO, Weitzberg E. Multiple single-breath measurements of nitric oxide in the intubated patient. *Am J Respir Crit Care Med Lung* 2003;168:1210-1215.
129. Ozkan M, Dweik RA, Laskowski D, Arroliga AC, Erzurum SC. High levels of nitric oxide in individuals with pulmonary hypertension receiving epoprostenol therapy. *Lung* 2001;179(4):233-243.
130. Humpl T, Campbell R, Stephens D, Van Arsdell G, Benson LN, Holtby HM, Slutsky AS, Adatia I. Levels of exhaled nitric oxide before and after surgical and transcatheter device closure of atrial septal defects in children. *J Thoracic Cardiovascular Surgery* 2002;124(4):806-810.
131. Cuthbertson BH, Stott SA, Webster NR. Exhaled nitric oxide as a marker of lung injury in coronary artery bypass surgery. *Br J Anaesthesiol* 2002;89(2):247-250.
132. Ali S, Metzger WJ, Olanrewaju HA, Mustafa SJ. Adenosine receptor-mediated relaxation of rabbit airway smooth muscle: a role for nitric oxide. *Am J Physiol* 1997;273(3 Pt 1):L581-L587.
133. Ermert M, Ruppert C, Gunther A, Duncker HR, Seeger W, Ermert L. Cell-specific nitric oxide synthase-isoenzyme expression and regulation in response to endotoxin in intact rat lungs. *Lab Invest* 2002;82(4):425-441.
134. Hunt JF, Fang K, Malik R, Snyder A, Malhotra N, Platts-Mills TA, Gaston B. Endogenous airway acidification. Implications for asthma pathophysiology. *Am J Respir Crit Care Med* 2000;161(3 Pt 1):694-699.
135. Yoshizumi M, Perrella MA, Burnett, JC Jr, Lee ME. Tumor necrosis factor downregulates an endothelial nitric oxide synthase mRNA by shortening its half-life. *Circ Res* 1993;73(1):205-209.
136. Marczin N, Antonov A, Papapetropoulos A, Munn DH, Virmani R, Kolodgie FD, Gerrity R, Catravas JD. Monocyte-induced downregulation of nitric oxide synthase in cultured aortic endothelial cells. *Arterioscler Thromb Vasc Biol* 1996;16(9):1095-1103.
137. Marczin N, Papapetropoulos A, Jilling T, Catravas JD. Prevention of nitric oxide synthase induction in vascular smooth muscle cells by microtubule depolymerizing agents. *Br J Pharmacol* 1993;109(3):603-605.
138. Stitt JT, Dubois AB, Douglas JS, Shimada SG. Exhalation of gaseous nitric oxide by rats in response to endotoxin and its absorption by the lungs. *J Appl Physiol* 1997;82(1):305-316.
139. Stewart TE, Valenza F, Ribeiro SP, Wener AD, Volgyesi G, Mullen JB, Slutsky AS. Increased nitric oxide in exhaled gas as an early marker of lung inflammation in a model of sepsis. *Am J Respir Crit Care Med* 1995;151(3 Pt 1):713-718.
140. Mehta S, Javeshghani D, Datta P, Levy RD, Magder S. Porcine endotoxemic shock is associated with increased expired nitric oxide. *Crit Care Med* 1999;27(2):385-393.
141. Vandivier RW, Eidsath A, Banks SM, Preas HL, Leighton SB, Godin PJ, Suffredini AF, Danner RL. Down-regulation of nitric oxide production by ibuprofen in human volunteers. *J Pharmacol Exp Ther* 1999;289(3):1398-1403.
142. Pedoto A, Caruso JE, Nandi J, Oler A, Hoffmann SP, Tassiopoulos AK, McGraw DJ, Camporesi EM, Hakim TS. Acidosis stimulates nitric oxide production and lung damage in rats. *Am J Respir Crit Care Med* 1999;159(2):397-402.
143. Tassiopoulos AK, Carlin RE, Gao Y, Pedoto A, Finck CM, Landas SK, Tice DG, Marx W, Hakim TS, McGraw DJ. Role of nitric oxide and

- tumor necrosis factor on lung injury caused by ischemia/reperfusion of the lower extremities. *J Vasc Surg* 1997;26(4):647-656.
144. Brett SJ, Evans TW. Measurement of endogenous nitric oxide in the lungs of patients with the acute respiratory distress syndrome. *Am J Respir Crit Care Med* 1998;157(3 Pt 1):993-997.
 145. Sittipunt C, Steinberg KP, Ruzinski JT, Myles C, Zhu S, Goodman RB, Hudson LD, Matalon S, Martin TR. Nitric oxide and nitrotyrosine in the lungs of patients with acute respiratory distress syndrome. *Am J Respir Crit Care Med* 2001;163(2):503-510.
 146. Brett SJ, Quinlan GJ, Mitchell J, Pepper JR, Evans TW. Production of nitric oxide during surgery involving cardiopulmonary bypass. *Crit Care Med* 1998;26(2):272-278.
 147. Kovesi T, Royston D, Yacoub M, Marczin N. Basal and nitroglycerin-induced exhaled nitric oxide before and after cardiac surgery with cardiopulmonary bypass. *Br J Anaesthesiol* 2003;90(5):608-616.
 148. Hill GE, Snider S, Galbraith TA, Forst S, Robbins RA. Glucocorticoid reduction of bronchial epithelial inflammation during cardiopulmonary bypass. *Am J Respir Crit Care Med* 1995;152(6 Pt 1):1791-1795.
 149. Marczin N, Riedel B, Gal J, Polak J, Yacoub M. Exhaled nitric oxide during lung transplantation. *Lancet* 1997;350(9092):1681-1682.
 150. Hunt J, Byrns RE, Ignarro LJ, Gaston B. Condensed expirate nitrite as a home marker for acute asthma [letter]. *Lancet* 1995;346:1235-1236.
 151. Redington AE, Meng QH, Springall DR, Evans TJ, Creminon C, Maclouf J, Holgate ST, Howarth PH, Polak JM. Increased expression of inducible nitric oxide synthase and cyclo-oxygenase-2 in the airway epithelium of asthmatic subjects and regulation by corticosteroid treatment. *Thorax* 2001;56:351-357.
 152. Hamid Q, Springall DR, Riveros-Moreno V, Chanez P, Howarth P, Redington A, Bousquet J, Godard P, Holgate S, Polak JM. Induction of nitric oxide synthase in asthma. *Lancet* 1993;342(8886-8887):1510-3.
 153. Stamler JS, Singel DJ, Loscalzo J. Biochemistry of nitric oxide and its redox-activated forms. *Science* 1992;258:1898-1902.
 154. Wu W, Chen Y, Hazen SL. Eosinophil peroxidase nitrates protein tyrosyl residues: implications for oxidative damage by nitrating intermediates in eosinophilic inflammatory disorders. *J Biol Chem* 1999;274:25933-25944.
 155. Weitzberg E, Lundberg JO. Nonenzymatic nitric oxide production in humans. *Nitric Oxide* 1998;2:1-7.
 156. Snyder AH, McPherson ME, Hunt JF, Johnson M, Stamler JS, Gaston B. Acute effects of aerosolized S-nitrosoglutathione in cystic fibrosis. *Am J Respir Crit Care Med* 2002;165:922-926.
 157. Gaston B, Ratjen F, Vaughan JW, Malhotra NR, Canady RG, Snyder AH, Hunt JF, Gaertig S, Goldberg JB. Nitrogen redox balance in the cystic fibrosis airway: effects of antipseudomonal therapy. *Am J Respir Crit Care Med* 2002;165:387-390.
 158. Latzin P, Griese M. Exhaled hydrogen peroxide, nitrite and nitric oxide in healthy children: decrease of hydrogen peroxide by atmospheric nitric oxide. *Eur J Med Res* 2002;7:353-358.
 159. Gessner C, Hammerschmidt S, Kuhn H, Lange T, Engelmann L, Schauer J, Wirtz H. Exhaled breath condensate nitrite and its relation to tidal volume in acute lung injury. *Chest* 2003;124:1046-1052.
 160. Machado RF, Londhe Nerkar MV, Dweik RA, Hammel J, Janocha A, Pyle J, Laskowski D, Jennings C, Arroliga AC, Erzurum SC. Nitric oxide and pulmonary arterial pressures in pulmonary hypertension. *Free Radic Biol Med* 2004;37:1010-1017.
 161. Kostikas K, Papatheodorou G, Ganas K, Psathakis K, Panagou P, Loukides S. pH in expired breath condensate of patients with inflammatory airway diseases. *Am J Respir Crit Care Med* 2002;165:1364-1370.
 162. Hunt JF, Fang K, Malik R, Snyder A, Malhotra N, Platts-Mills TA, Gaston B. Endogenous airway acidification: implications for asthma pathophysiology. *Am J Respir Crit Care Med* 2000;161:694-699.
 163. Tate S, MacGregor G, Davis M, Innes JA, Greening AP. Airways in cystic fibrosis are acidified: detection by exhaled breath condensate. *Thorax* 2002;57:926-929.
 164. Gessner C, Kuhn H, Engelmann L, Schauer J, Wirtz H. Airway acidification in artificial ventilated patients. *Eur Respir J* 2001;18:480s.
 165. Vaughan J, Ngamtrakulpanit L, Pajewski T, Turner R, Nguyen TA, Smith A, Urban P, Hom S, Gaston B, Hunt J. Exhaled breath condensate pH is a robust and reproducible assay of airway acidity. *Eur Respir J* 2003;22:889-894.
 166. Hunt JF, Erwin E, Palmer L, Vaughan J, Malhotra N, Platts-Mills TAE. Expression and activity of pH-regulatory glutaminase in the human airway epithelium. *Am J Respir Crit Care Med* 2002;165:101-107.
 167. Effros RM. Do low exhaled condensate NH₄⁺ concentrations in asthma reflect reduced pulmonary production? *Am J Respir Crit Care Med* 2003;167(1):91; author reply 91-92.
 168. Wells K, Vaughan J, Pajewski TN, Hom S, Ngamtrakulpanit L, Smith A, Nguyen A, Turner R, Hunt JF. Exhaled breath condensate pH assays are not influenced by oral ammonia. *Thorax* 2005;60:27-31.
 169. Effros RM, Hoagland KW, Bosbous M, Castillo D, Foss B, Dunning M, Gare M, Lin W, Sun F. Dilution of respiratory solutes in exhaled condensates. *Am J Respir Crit Care Med* 2002;165:663-669.
 170. Vaughan J, Ngamtrakulpanit L, Pajewski T, Turner R, Nguyen TA, Smith A, Urban P, Hom S, Gaston B, Hunt J. Exhaled breath condensate pH is a robust and reproducible assay of airway acidity. *Eur Respir J* 2003;22:889-894.
 171. Lee KH, Rico P, Billiar TR, Pinsky MR. Nitric oxide production after acute, unilateral hydrochloric acid-induced lung injury in a canine model. *Crit Care Med* 1998;26:2042-2047.
 172. Gaston B, Sears S, Woods J, Hunt J, Ponaman M, McMahon T, Stamler JS. Bronchodilator S-nitrosothiol deficiency in asthmatic respiratory failure. *Lancet* 1998;351:1317-1319.
 173. Ojoo JC, Mulrennan A, Kastelik JA, Morice AH, Redington AE. Exhaled breath condensate pH and exhaled nitric oxide in allergic asthma and in cystic fibrosis. *Thorax* 2005;60:22-26.
 174. Hunt J. The informative complexity of exhaled nitrogen oxide chemistry. *Thorax* 2005;60:2-3.
 175. Jobsis Q, Raatgeep HC, Schellekens SL, Hop WC, Hermans PW, de Jongste JC. Hydrogen peroxide in exhaled air of healthy children: reference values. *Eur Respir J* 1998;12:483-485.
 176. Schleiss MB, Holz O, Behnke M, Richter K, Magnussen H, Jorres RA. The concentration of hydrogen peroxide in exhaled air depends on expiratory flow rate. *Eur Respir J* 2000;16:1115-1118.
 177. De Benedetto F, Aceto A, Dragani B, Spacone A, Formisano S, Cocco R, Sanguinetti CM. Validation of a new technique to assess exhaled hydrogen peroxide: results from normals and COPD patients. *Monaldi Arch Chest Dis* 2000;55:185-188.
 178. Corradi M, Folesani G, Andreoli R, Manini P, Bodini A, Piacentini G, Carraro S, Zanconato S, Baraldi E. Aldehydes and glutathione in exhaled breath condensate of children with asthma exacerbation. *Am J Respir Crit Care Med* 2003;167:395-399.
 179. van Beurden WJ, Harff GA, Dekhuijzen PN, van den Bosch MJ, Creemers JP, Smeenk FW. An efficient and reproducible method for measuring hydrogen peroxide in exhaled breath condensate. *Respir Med* 2002;96:197-203.