## ATS Workshop Proceedings: Exhaled Nitric Oxide and Nitric Oxide Oxidative Metabolism in Exhaled Breath Condensate

## **Executive Summary**

This official executive summary of an American Thoracic Society Workshop Report was approved by the ATS Board of Directors, September 2005.

## **BACKGROUND TO WORKSHOP**

The field of noninvasive assessment of airway inflammation has developed rapidly since the discoveries in the late 1980s that nitric oxide (NO) is a central biological mediator and, in 1991, that exhaled breath contained NO (1). Accompanying this was the realization that asthma is an inflammatory airway syndrome, best treated by antiinflammatory agents. Subsequently, exhaled NO was found to be high in asthma (2), and to fall after antiinflammatory drugs but not bronchodilators. Today, exhaled NO (FE<sub>NO</sub>) is progressing from a research tool to clinical application in asthma (3). In the mid-1990s, the field of exhaled breath condensate (EBC) analysis, a biological sample of the airways took off rapidly, opening up another way to sample airway chemistry in a noninvasive manner. Exhaled NO measurement and NO redox assessment in EBC were the focus of an American Thoracic Society (ATS) workshop that was held in Toronto in 2002 and was divided into two sections: (1) exhaled and nasal NO and (2) lung nitrogen oxide and redox assessment using EBC. The full workshop proceedings can be accessed in the current proceedings of the ATS.

## **EXHALED NO**

The workshop commenced with updates on the measurement of  $F_{E_{NO}}$  since the previous 1998 ATS workshop, which resulted in the 1999 ATS statement on exhaled and nasal NO measurement (4). Several new sensor technologies to detect NO were presented, and experience with the 1999 online and offline exhaled NO measurement recommendations was reviewed. In general, the methods recommended in 1999 had met with success and acceptance, although several minor modifications were adopted.

For online  $F_{E_{NO}}$ , the need to inspire to total lung capacity (TLC) was relaxed to a near-TLC inspiration for patients who could not inspire fully. Also, as  $F_{E_{NO}}$  is so reproducible, it was believed that two reproducible measures were adequate instead of the previous requirement for three reproducible measures. The online flow rate of 50 ml/s was believed to be acceptable and reproducible for both adults and children. The section on  $F_{E_{NO}}$  plateau definition was revised to make defining this plateau clearer.

For offline exhaled NO methods, there was discussion around the 1999 flow rate of 350 ml/s, which is much higher than the online method (50 ml/s) (4). It was agreed, however, that investi-

Am J Respir Crit Care Med Vol 173. pp 811–813, 2006 DOI: 10.1164/rccm.2601014 Internet address: www.atsjournals.org gators could use other flow rates for offline measurement as long as this is reported in manuscripts. Pediatric exhaled NO measurement was reviewed only briefly as there is a recent ATS/ European Respiratory Society (ERS) task force that published recommendations in 2002 (5).

Clinical application of exhaled NO was thought to be limited to asthma at this stage, and the evidence for the use of this marker in asthma was reviewed.

Techniques of nasal NO measurement were discussed, with a general consensus that the 1999 nasal NO recommendations were adequate for most research purposes (4). Regarding the clinical application of nasal NO, it was decided that the only promising application was in screening for primary ciliary dyskinesia syndromes (6), and that a simple widely applicable method for this application should be developed.

The development of NO excretion models for the lung with a description of flow-independent NO exchange parameters was reviewed (7). Finally, approaches to NO measurement in the ventilated patient, such as the online single-breath method (8), were presented and discussed by a separate breakout group.

One purpose of the workshop was to revise and update the 1999 ATS exhaled and nasal NO recommendations. This was achieved at a final session and the 2005 ATS/ERS statement on exhaled and nasal NO measurement has now appeared in the *AJRCCM* (9), and can be accessed from the ERS website and was accompanied by an editorial (10).

# LUNG NITROGEN OXIDE AND REDOX ASSESSMENT USING EBC

## Background

This workshop focused on the potential of EBC to provide information regarding lung nitrogen and oxygen redox balance. Methods by which measurements of higher oxides of nitrogen (e.g., nitrite  $[NO_2^- \text{ and } NO_3^-]$ ) in EBC, as well as oxidants such as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), and acidity that could complement the more established exhaled NO test were presented.

## Anatomic Source of EBC Components

Oral collections of EBC are the most commonly used, although nasal and endotracheal collections are increasingly reported. All nonvolatile compounds found in EBC come from either the droplets/particles evolved from the airway lining fluid (ALF; oropharynx, trachea, bronchi, alveolae) or are reaction products of volatiles that enter EBC from gas phase. Although endotracheal collections of EBC exclude oropharyngeal contribution, most studies of EBC involve oral collections. Even a small contribution of saliva to an EBC sample may be relevant, as the concentrations in EBC of nonvolatile substances are generally very low (micromolar range or below). There are as yet no data regarding the extent to which aerosolization of alveolar fluid

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contributes to EBC. Because alveolar fluid moves toward the proximal airways, there is likely to be some alveolar representation in EBC. Indeed, the first studies of EBC involved surfactant (11).

#### Nitrogen Oxides

Deviations of EBC nitrogen oxide (NOx) concentrations, or ratios among them, may reflect the oxidative and NOx conditions of the airway, including not just formation of NO, but all the various inorganic, eukaryotic, and prokaryotic reactions that occur around NO (12). These assays may then reflect NO synthase (NOS) activity, airway inflammation, innate immune responses, bacterial burdens, airway pH, and nitrergic neurotransmission. Importantly, assays for the higher oxides of nitrogen complement knowledge gained from exhaled NO assays. For example, although the cystic fibrosis lung has prominent inflammation, exhaled NO levels are generally low. However EBC NOx are found at high concentrations (13), perhaps suggesting that the NO, although indeed being produced in larger amounts, is being oxidized. Another appeal of EBC NOx is less intuitive, but intriguing, as data support that EBC NO<sub>2</sub><sup>-</sup> levels provide knowledge regarding lung overexpansion in intubated subjects (14).

#### EBC pH Assays

Only recently has progress been made in understanding the role of airway pH in respiratory illness. One theoretic underpinning for studying EBC pH is that volatile airway acids could be trapped in EBC, and that they would be exhaled to a greater extent from an acidic source fluid. EBC pH has been found to be low in many respiratory diseases, and increased attention has been given to airway acid-base balance. EBC pH assays have undergone more extensive validation experiments than many other assays in EBC. Although with oral collections EBC will always in part represent (or be contaminated by) the oropharynx, a relevant contribution of orally derived ammonia to the EBC pH has not been identified in specific experiments. Gas standardization or deaeration of EBC prior to measurement of pH provides different information than measurement of pH immediately after collection. Both methodologies may have advantages. Gastroesophageal reflux of acidic fluid with or without microaspiration into the trachea is one potential mechanism of EBC acidification that has been considered, and is under active investigation in several laboratories (15). Alternate explanations for EBC acidification include lower airway acid production by numerous described pathways. In all likelihood, any process that acidifies any level of the airway could lead to EBC acidification if anions present in the fluid become protonated and volatilized.

#### Interactions of pH and NOx

NO produced in the lower airway may be exhaled but could also undergo chemical change, including oxidation to NOx. As noted, these nonvolatile NOx can be exhaled and serve as a marker of NOS activity that complements and expands on the information provided by exhaled NO. Experimental acidification of the ALF is known to lead to rapid and substantial increases in exhaled NO (16), likely through protonation and breakdown of NO<sub>2</sub><sup>-</sup>. However, an association between EBC pH and exhaled NO is not generally identified in clinical studies (13). Acid and NOx together increase the likelihood of formation of S-nitrosothiols as well as nitrated proteins, which affect diverse aspects of lung physiology and immunology.

#### Hydrogen Peroxide

EBC has provided the strongest evidence to date of oxidative disturbance in lung diseases. With more supportive validation

studies than most other EBC markers,  $H_2O_2$  has received extensive attention (17). The flow dependence of EBC  $H_2O_2$  suggests that it arrives in EBC partially in the gas phase, and may arise from the airway wall, similar to exhaled NO (18). The reactivity of  $H_2O_2$  makes it necessary to handle specimens expeditiously.

# Evidence of Oxidation by Measurement of Larger Molecules in EBC

Larger molecules exist in EBC (e.g, lipids). Evidence of lipid oxidation can be derived from the presence of such substances as malonaldehyde and 8-isoprostane, which have been found to be particularly informative in diverse respiratory and nonrespiratory disease states. Glutathione, both in reduced and oxidized form, has been identified in EBC and the levels or ratios of these compounds are particularly interesting in assessing the overall redox balance of the lung more comprehensively (19).

Still clouded by uncertainties regarding anatomic sources and complex chemistries, EBC nonetheless allows us to gain cautious insights into ALF constituents in disease states and in health. Lung redox information has become obtainable through this technology, through assays of pH, nitrogen oxides, hydrogen peroxide, glutathione, and markers of the presence of disturbances in these compounds, such as nitrotyrosine, isoprostane, and aldehydes. EBC has potential to complement  $FE_{NO}$  to more fully elucidate the nitrogen oxide chemistry of the airways and lung. As a marker of disease activity, EBC assays may monitor acid stress, oxidative stress, and inflammation, separate but often-related entities to which many of our therapies are directed.

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**Conflict of Interest Statement:** P.E.S. is currently a full-time employee of AstraZeneca but this bears no relationship or conflict of interest to this document; previously, he was a paid consultant for lonics-Sievers Instruments and Aerocrine AB, manufacturers of exhaled NO meters; he receives royalties from Ionics-Sievers Instruments and Aperon Biosystems for licensed patents. J.H. is a cofounder and substantial shareholder of Respiratory Research, Inc., which designs and manufactures exhaled breath condensate collection equipment and has licensed exhaled breath condensate pH and other assays from the University of Virginia; he is an inventor of exhaled breath condensate nitrogen oxide and pH assays.

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#### American Thoracic Society Documents

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#### References

- Gustafsson LE, Leone AM, Persson MG, Wiklund NP, Moncada S. Endogenous nitric oxide is present in the exhaled air of rabbits, guinea pigs and humans. *Biochem Biophys Res Commun* 1991;181:852–857.
- Alving K, Weitzberg E, Lundberg JM. Increased amount of nitric oxide in exhaled air of asthmatics. *Eur Respir J* 1993;6:1368–1370.
- Bates CA, Silkoff PE. Exhaled nitric oxide in asthma: from bench to bedside. J Allergy Clin Immunol 2003;111:256–262.
- American Thoracic Society. Recommendations for standardized procedures for the on-line and off-line measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide in adults and children-1999. *Am J Respir Crit Care Med* 1999;160:2104–2117.
- Baraldi E, de Jongste JC. Measurement of exhaled nitric oxide in children, 2001. Eur Respir J 2002;20:223–237.
- Bush A, Cole P, Hariri M, Mackay I, Phillips G, O'Callaghan C, Wilson R, Warner JO. Primary ciliary dyskinesia: diagnosis and standards of care. *Eur Respir J* 1998;12:982–988.
- George SC, Hogman M, Permutt S, Silkoff PE. Modeling pulmonary nitric oxide exchange. J Appl Physiol 2004;96:831–839.

- Tornberg DC, Bjorne H, Lundberg JO, Weitzberg E. Multiple singlebreath measurements of nitric oxide in the intubated patient. *Am J Respir Crit Care Med* 2003;168:1210–1215.
- American Thoracic Society/European Respiratory Society. ATS/ERS recommendations for standardized procedures for the online and offline measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide, 2005. Am J Respir Crit Care Med 2005;171:912–930.
- Silkoff PE. Monitoring nitric oxide: here to stay for bench and bedside. Eur Respir J 2005;25:949–950.
- Sidorenko GI, Zborovskii EI, Levina DI. Surface-active properties of the exhaled air condensate (a new method of studying lung function). *Ter Arkh* 1980;52:65–68.
- Gaston B, Ratjen F, Vaughan JW, Malhotra NR, Canady RG, Snyder AH, Hunt JF, Gaertig S, Goldberg JB. Nitrogen redox balance in the cystic fibrosis airway: effects of antipseudomonal therapy. *Am J Respir Crit Care Med* 2002;165:387–390.
- Ojoo JC, Mulrennan SA, Kastelik JA, Morice AH, Redington AE. Exhaled breath condensate pH and exhaled nitric oxide in allergic asthma and in cystic fibrosis. *Thorax* 2005;60:22–26.
- Gessner C, Hammerschmidt S, Kuhn H, Lange T, Engelmann L, Schauer J, Wirtz H. Exhaled breath condensate nitrite and its relation to tidal volume in acute lung injury. *Chest* 2003;124:1046–1052.
- Ricciardolo FL, Gaston B, Hunt J. Acid stress in the pathology of asthma. J Allergy Clin Immunol 2004;113:610–619.
- Lee KH, Rico P, Billiar TR, Pinsky MR. Nitric oxide production after acute, unilateral hydrochloric acid-induced lung injury in a canine model. *Crit Care Med* 1998;26:2042–2047.
- Jobsis Q, Raatgeep HC, Schellekens SL, Hop WC, Hermans PW, de Jongste JC. Hydrogen peroxide in exhaled air of healthy children: reference values. *Eur Respir J* 1998;12:483–485.
- Schleiss MB, Holz O, Behnke M, Richter K, Magnussen H, Jorres RA. The concentration of hydrogen peroxide in exhaled air depends on expiratory flow rate. *Eur Respir J* 2000;16:1115–1118.
- Corradi M, Folesani G, Andreoli R, Manini P, Bodini A, Piacentini G, Carraro S, Zanconato S, Baraldi E. Aldehydes and glutathione in exhaled breath condensate of children with asthma exacerbation. *Am J Respir Crit Care Med* 2003;167:395–399.