COVID-19 ACUTE MYOCARDIAL INJURY

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RETROSPECTIVE CLINICAL TRIALS

- COVID-19 primarily effects the upper respiratory tract causing pneumonia, respiratory failure and acute respiratory distress syndrome, there have also been many reports of cardiovascular involvement
 - Retrospective Single Study Trials
 - Huang et al. Lancet 2020
 - Chen et al. Lancet 2020
 - Wang et al. JAMA 2020
 - Retrospective Multi Center Studies
 - Wu et al. JAMA 2020
 - Guan et al. NEJM 2020
- COVID-19 infection can also present with isolated cardiac symptoms, even in the absence of respiratory symptoms (Inciardi *et al. JAMA Cardiol* 2020)

JAMA Cardiology | Brief Report

Cardiac Involvement in a Patient With Coronavirus Disease 2019 (COVID-19)

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- 53F with no prior medical history presenting to Niguarda Hospital in Milan, Italy in March 2020 with chest pain and dyspnea
- Presenting VS: afebrile, HR 100 bpm, BP 90/50 mmHg, SpO2 98% RA

Figure 1. Electrocardiographic and Chest Radiographic Findings

A Electrocardiography



A, Electrocardiography showing sinus rhythm with low voltage in the limb leads, Posteroanterior chest radiography at presentation. No thoracic abnormalities diffuse ST-segment elevation (especially in the inferior and lateral leads), and were noted. ST-segment depression with T-wave inversion in leads V1 and aVR. B,

Table. Clinical Laboratory Results								
		Result						
Measure	Reference range	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7
Red blood cell count, ×10 ⁶ /µL	4.0-5.2	5.5ª	4.6	4.0 ^b	3.9 ^b	3.8 ^b	3.6 ^b	3.7 ^b
Hemoglobin, g/dL	12.0-16.0	17.1ª	14.5	12.4	11.9 ^b	12.0	11.4 ^b	11.2 ^b
Hematocrit, %	37.0-47.0	49.3ª	42.1	36.0 ^b	34.9 ^b	35.1 ^b	33.9 ^b	33.6 ^b
White blood cell count, per µL	4000-10 800	8900	12 090ª	9920	10 900	13 470ª	13 730 ^a	13 500ª
Lymphocyte count								
Relative, %	20.0-40.0	10.6 ^b	NA	NA	NA	NA	NA	7.7 ^b
Absolute, per µL	900-4000	950	NA	NA	NA	NA	NA	1040
Platelet count, ×10 ³ /µL	130-400	152	168	164	213	317	317	360
Sodium, mEq/L	136-145	129 ^b	133 ^b	129 ^b	136	132 ^b	134 ^b	137
Potassium, mEq/L	3.4-4.5	5.7ª	6.3ª	3.9	3.7	3.5	3.6	3.6
Chloride, mEq/L	98-107	89 ^b	96 ^b	92 ^b	92 ^b	NA	92 ^b	94 ^b
Calcium, mg/dL	8.60-10.20	8.63	NA	7.84 ^b	8.15 ^b	NA	NA	NA
Creatinine, mg/dL	0.60-1.00	0.75	0.76	0.53 ^b	0.88	0.99	0.96	0.80
C-reactive protein, mg/dL	<0.5	1.3ª	0.7 ^a	1.0 ^a	1.1ª	0.6	0.4	0.3
Creatine kinase-MB, ng/mL	<4.9	20.3ª	39.9ª	30.7ª	13.3	5.2	3.3	2.8
High-sensitivity troponin T, ng/mL	<0.01	0.24	0.59	0.78	0.89	0.76	0.65 ^a	0.63ª
NT-proBNP, pg/mL	<300 ^c	5647	8465	8133	5113	2827	NA	NA

Figure 2. 1.5-Tesla Cardiac Magnetic Resonance Imaging

A STIR sequence in short-axis view

B STIR sequence in 4-chamber view



C T2-mapping sequence in short-axis view

D T2-mapping sequence in 4-chamber view





E PSIR sequence in short-axis view

F PSIR sequence in 4-chamber view



RETROSPECTIVE CLINICAL TRIALS

- Retrospective studies from Wuhan University examining cardiovascular disease in COVID-19 (Guo et al. JAMA Cardiol 2020, Shi et al. JAMA Cardiol 2020)
 - Patients with baseline cardiovascular disease have increased mortality during COVID-19
 - 7.62% mortality in patients without prior CVD and with normal TnT
 - 13.33% morality in patients WITH prior CVD and with normal TnT
 - Patients who experience acute myocardial injury during COVID-19 infection have worse mortality even in the absence of baseline symptoms (although baseline cardiovascular disease + acute myocardial injury had higher mortality)
 - 37.5% mortality in patients without prior CVD with ELEVATED TnT
 - 69.44% mortality in patients WITH prior CVD and with ELEVATED TnT
 - Acute myocardial injury alone, even without LV dysfunction, was associated with higher mortality, however those with LV dysfunction had the worst mortality of any age group
- Cardiovascular complications of COVID-19 infection are a major contributor to patient mortality, but the pathophysiology underlying this cardiac injury is not presently understood

PROPOSED MECHANISMS OF MYOCARDIAL INJURY

- Type I MI/Plaque Rupture
 - Increased rates of type I MI in influenza (Nguyen JAMA Cardiol 2016, Kwong NEJM 2018)
- Type II MI/Demand Ischemia
 - Similar to that seen in severe sepsis
- Acute Fulminant Myocarditis
 - Similar to that seen with MERS (Alhogbani Ann Saudi Med 2016)
 - Would require viremia and direct infection of myocardium since viral entry is most likely mediated by infection of nasopharyngeal cells, and virus was detected in blood in only a minority of patients (To Lancet Infect Dis 2020)
- Cytokine Storm-mediated Injury
 - Autoimmune response to viral infection mediates end-organ damage
 - "Secondary hemophagocytic lymphohistiocytosis"
- ACE2-mediated direct infection of myocardial cells (Oudit J Clin Invest 2009, Wrapp Science 2020, Patel Circ Res 2016)
 - Direct infection of cardiomyocytes
 - Vascular/Endothelial dysfunction

• Limited myocardial tissue pathology has been completed to date

Bonow, Fonarow, O'Gara, Yancy. JAMA Cardiology 2020



- 69M presents to ED in Lombardy, Italy with cough, shortness of breath and weakness x 4 days
- CT Thorax with bilateral interstitial infiltrates, labs with leukocytosis and elevated inflammatory markers, ABG with pH 7.2
- TTE with LVEF 35% \rightarrow 25% within 3 hours
- Cath unremarkable → IABP → worsening hypotension → VA-ECMO + intubation
- Transfer to tertiary MC \rightarrow EMB performed

Myocardial localization of coronavirus in COVID-19 cardiogenic shock

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Figure 1 Light microscopy immunostaining of the inflammatory infiltrate. (A,B) Low- and high-power views of endomyocardial biopsy, with sparse CD45RO positive interstitial cells. (C,D) Large, vacuolated macrophages immunostained with anti-CD68 antibodies. (E) Ultrastructural morphology of a large and cytopathic macrophage. (A–D: the bar scale is in the left low corner of each panel. E: the bar scale is in the right low corner of the panel and corresponds to 2 µm).



Figure 2 Examples of small groups of viral particles (A and B; panel C shows a higher magnification of one of the viral particles squared in dashed red box of panel B) or single particles (D-F) observed within the interstitial cells of the myocardium of the patient. The red arrows indicate the most typical and easy-to-recognize viral particles, whose size varies from about 70 nm to 120 nm (see the white bars in the panels). Morphology also shows small differences with more or less prominent spikes of the viral crown. The morphology may also show viral particle disruption (E, green arrow) or attenuation of spikes of the crown (D and F), or viral particles in budding attitude (F). (Bar scale: A and B, 200 nm; C, 50 nm; D, 100 nm; F, 50 nm).

Bradley *et al.* Histopathological and Ultrastructural Findings in COVID-19 Infection <u>https://www.medrxiv.org/content/10.1101/2020.04.17</u>.

<u>20058545v1</u>

Pest mortem analysis of 12 fatal cases presenting in Seattle, WA Feb-Mar 2020 (University of Washington)



Heart with lymphocytic myocarditis and myocytolysis No other details about case were provided



Fox et al. Pulmonary and Cardiac Pathology in COVID-19: The First Autopsy Series from New Orleans https://www.medrxiv.org/content/10.1101/2020.04.06.2005057 5v1.full.pdf

Post mortem analysis of 4 fatal cases at University Medical Center in New Orleans, LO (LSU/Tulane)



H&E stains of cardiac myocytes with focal degeneration (blue arrows). Myocardium did not show any large or confluent areas of myocyte necrosis but did show scattered individual cell necrosis in each heart examined. They did note some lymphocytes adjacent to (but not surrounding) these individual necrotic myocytes.

Possibly early lymphocytic myocarditis

CONCLUSIONS

- Limited myocardial tissue pathology available
- Patient demographics from the autopsy series are limited
- No basic transcriptomic/molecular data available
- Limited cardiac functional data available